

# Consistency Principle In Biological Dynamical Systems

Kunihiko Kaneko

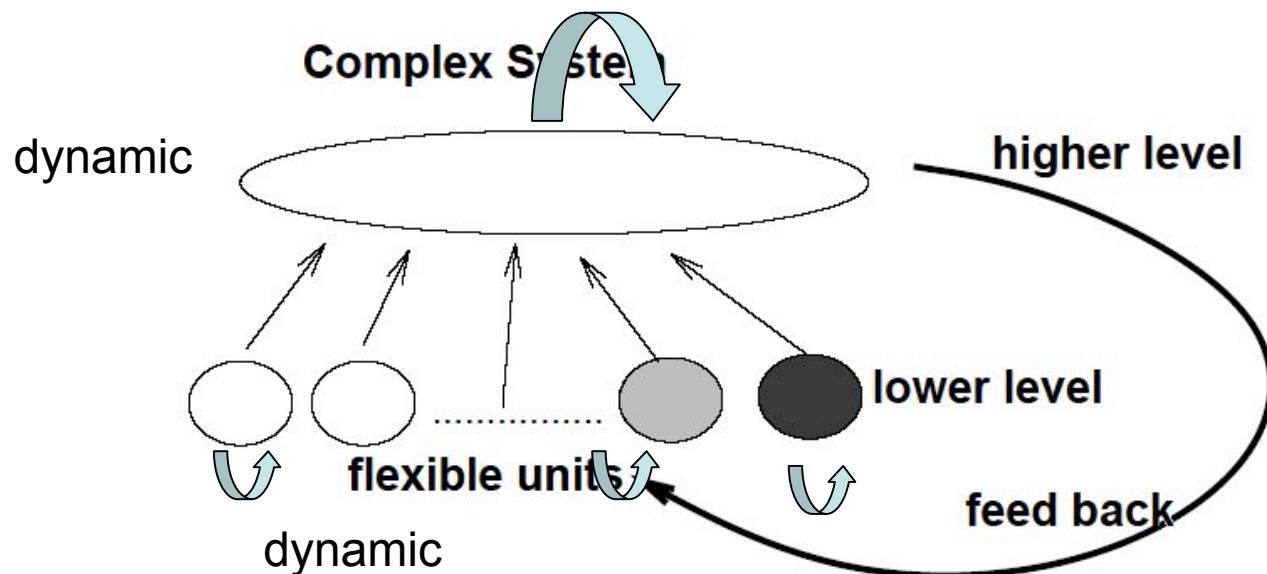
## Complex Systems Biology

Understand **Universal** features of Biological System

--Mutual dependence between parts and whole

**Guiding Principle:**

**Consistency between different levels**

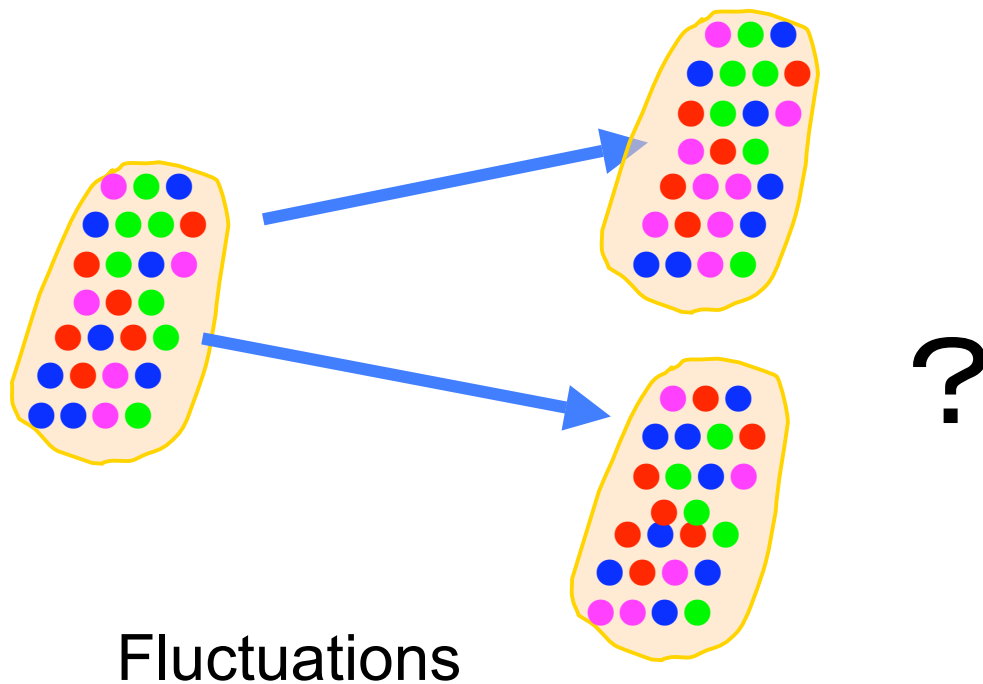


- **Cell reproduction vs molecule replication (briefly review) 15%**
  - **Genetic change (evolution) vs Phenotypic Fluctuation 75%**
  - \* **Gene expression vs Growth – Adaptation 5%**
  - \* **Reproduction of multicellular organism vs of cells (briefly) (development) 5%**
- 

*Underlying Biological Motivation;*

- \* Plasticity - Phenotypic Fluctuation- Evolvability
- \* Link between development and evolution
- \* Evolution of Robustness;
  - which type of systems is selected

How is recursive production of a cell sustained?  
each cell complex reaction network  
with diversity of chemicals;  
The number of molecules of each species not so  
large



# Toy Cell Model with Catalytic Reaction Network

## 'Crude but whole cell model'

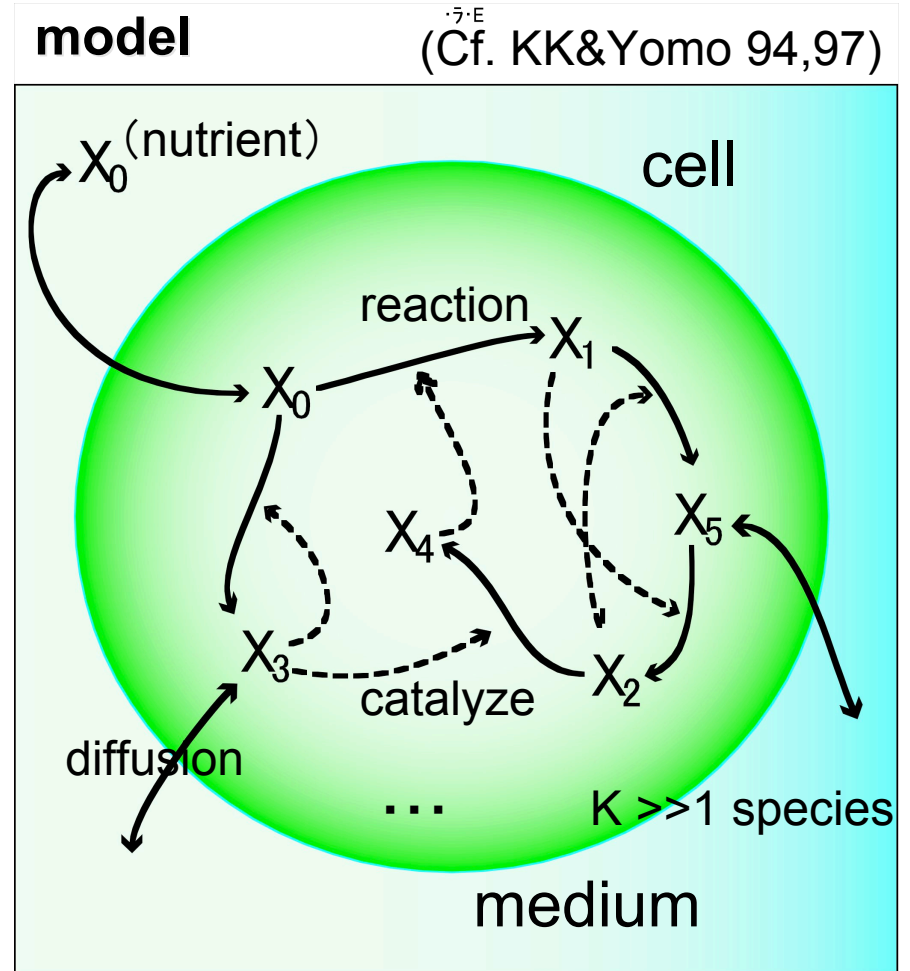
C.Furusawa & KK, PRL2003

■ **k species of chemicals** ,  $X_0 \cdots X_{k-1}$   
number ---  $n_0, n_1 \dots n_{k-1}$

■ **random catalytic reaction network**  
with the path rate  $p$   
for the reaction  $X_i + X_j \rightarrow X_k + X_l$

■ some chemicals are **penetrable**  
**through the membrane with the**  
**diffusion coefficient  $D$**

■ resource chemicals are thus  
transformed into impenetrable  
chemicals, leading to the growth in  
 $N = \sum n_i$  when it exceeds  $N_{\max}$   
**the cell divides into two**



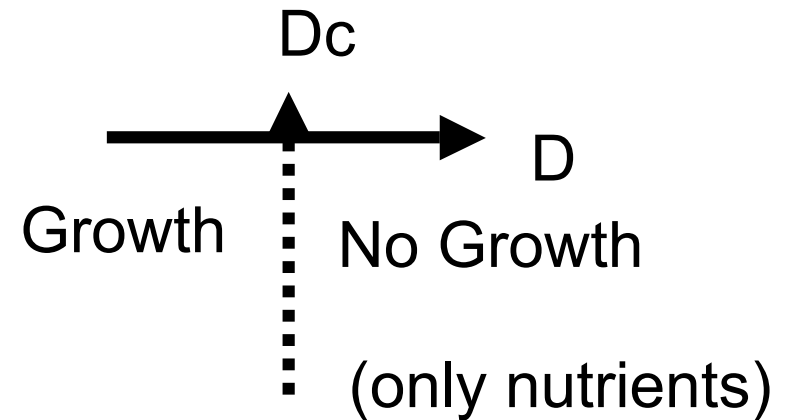
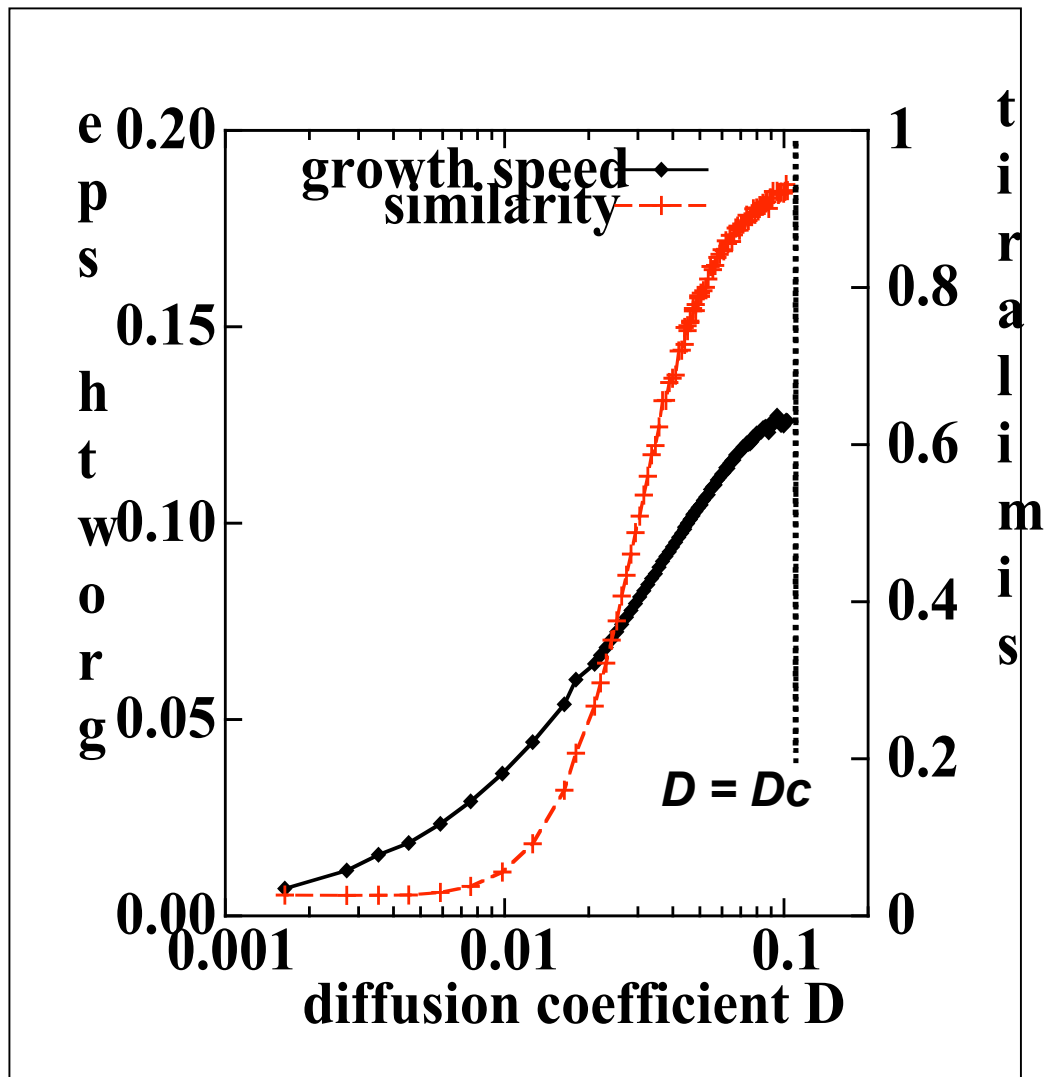
$dX_1/dt \propto X_0 X_4$ ; rate equation;  
Stochastic model here

In continuum description, the following rate eqn.,  
but we mostly use stochastic simulation

$$\begin{aligned} dn_i/dt = & \sum_{j,\ell} \text{Con}(j, i, \ell) \epsilon n_j n_\ell / N^2 \\ & - \sum_{j',\ell'} \text{Con}(i, j', \ell') \epsilon n_i n_{\ell'} / N^2 \\ & + D\sigma_i(\bar{n}_i/V - n_i/N), \end{aligned}$$

where  $\text{Con}(i, j, \ell)$  is 1 if there is a reaction  $i + \ell \rightarrow j + \ell$ , and 0 otherwise, whereas  $\sigma_i$  takes 1 if the chemical  $i$  is penetrable, and 0 otherwise. The third term describes the transport of chemicals through the membrane, where  $\bar{n}_i$  is

☆ Growth speed and fidelity in replication are maximum at  $D_c$

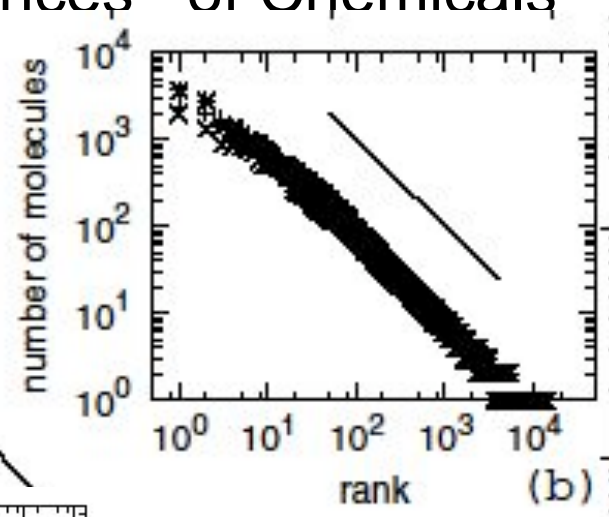
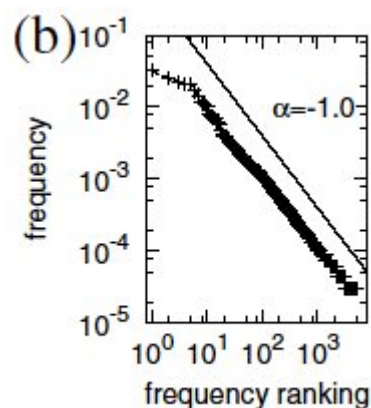
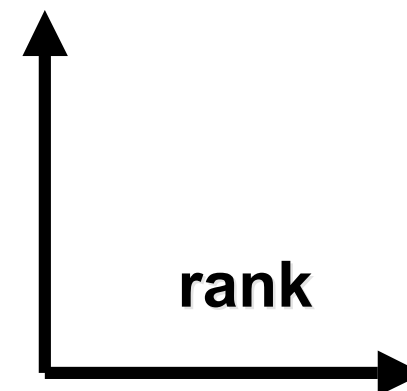


※similarity is defined from inner products of composition vectors between mother and daughter cells

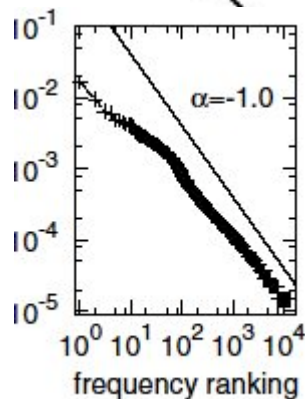
## Traces: universal statistics

## Power Law in Abundances of Chemicals

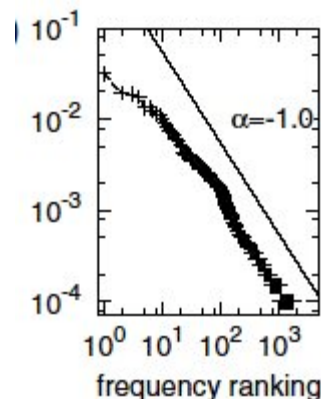
Theory;  
To keep reproduction  
preserving  
compositions →


 $n_i$  (number of molecules)


Human kidney, mouse ES



yeast



	number	rank
X1	300	5
X2	8000	1
X3	5000	2
X4	700	4
X5	2000	3
..... (for example)		

Average number of each chemical  $\propto 1/(\text{its rank})$

- Remarks:

- (0) **Universality**

- (1) **Evolution** to the critical state (with Zipf law) is confirmed numerically

- (2) Evolution to scale-free network appears later as embedding of power-law abundances into **topology of network** (Furusawa, KK, PRE 2006)

- (3) **Self-organization** to critical state, if transport of ‘nutrition chemicals’ is catalyzed by some chemicals (no need for choice of  $D$ ) (instead of simple diffusion) (Furusawa, KK, 2007)

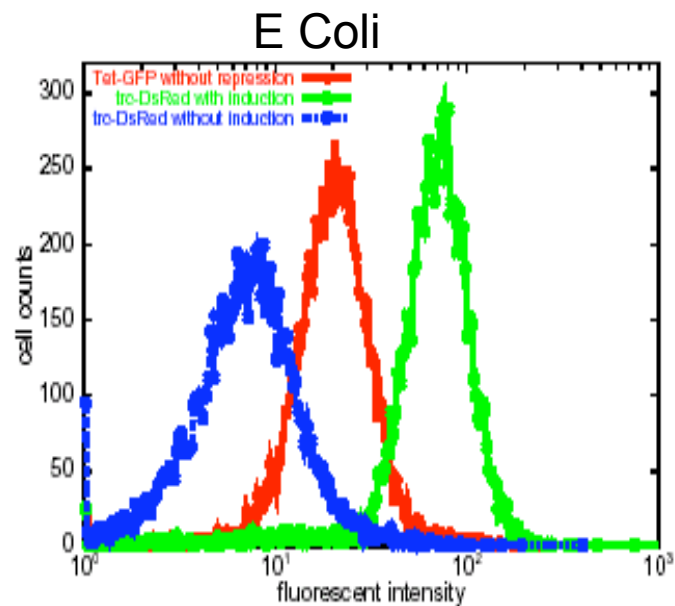


: fluctuation by cells:  
distribution of each Ni by cells

Furusawa,..  
KK,  
Biophysics2005

e.g.

cell1	X1	10000
cell2		8000
cell3		15000
cell4		20000



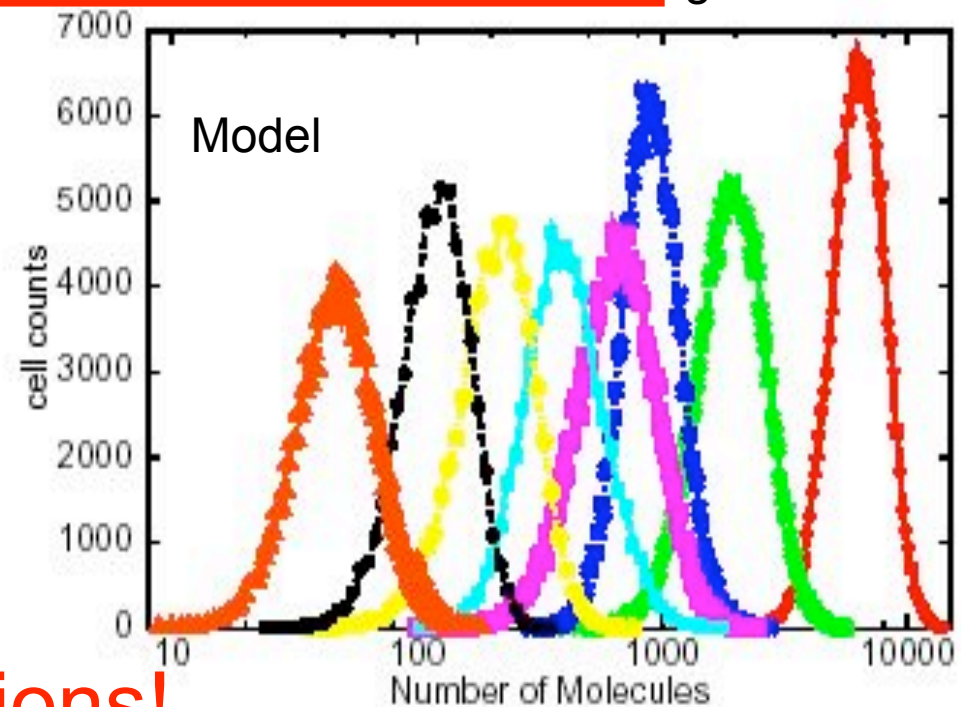
number distribution of the proteins measured by fluorescent intensity.  
from: Escherichia coli cell populations containing different reporter plasmids

Each color gives different  
chemical species

**QUITE LARGE Fluctuations!**

Log normal distribution!

histogram



**LOG SCALE**

number distribution of the molecules of chemical abundances. Distribution

## ☆Heuristic explanation of log-normal distribution

Consider the case that a component X is catalyzed by other component A, and replicate; the number -- $N_X$ 、 $N_A$

$$d N_X /dt = N_X N_A$$

then

$$d \log( N_X )/dt = N_A$$

If、 $N_A$  fluctuates around its mean  $\langle N_A \rangle$ , with fluct.  $\eta (t)$

$$d \log( N_X )/dt = \langle N_A \rangle + \eta (t)$$

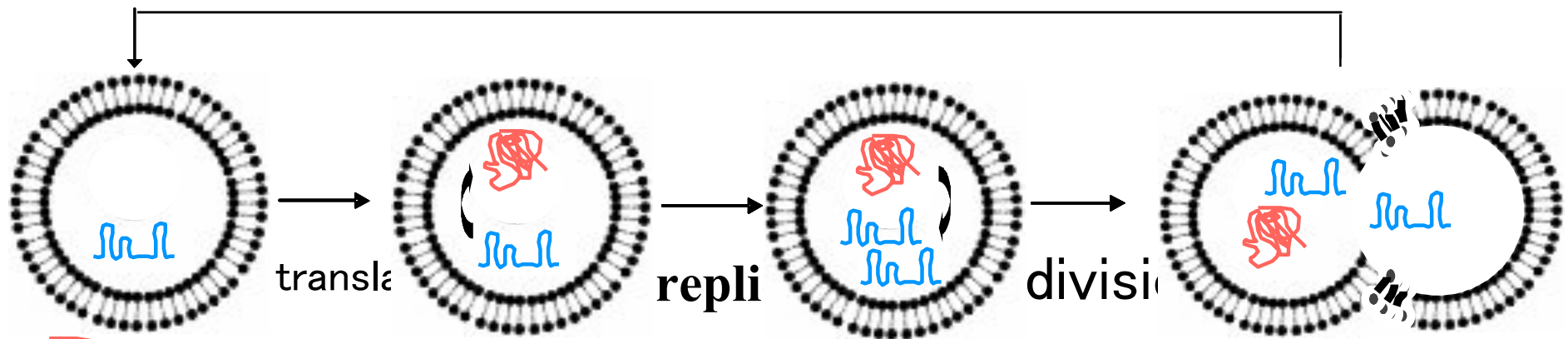
$\log( N_X )$  shows Brownian motion  $\rightarrow N_X$  log-normal distribution

too, simplified, since no direct self-replication exists here

But with cascade catalytic reactions, fluctuations are successively multiplied, (cf addition in central limit theorem.);Hence after logarithm, central limit th. applied

# Replicating artificial cell (experiment)

( $\leftrightarrow$  consistency, minority control)

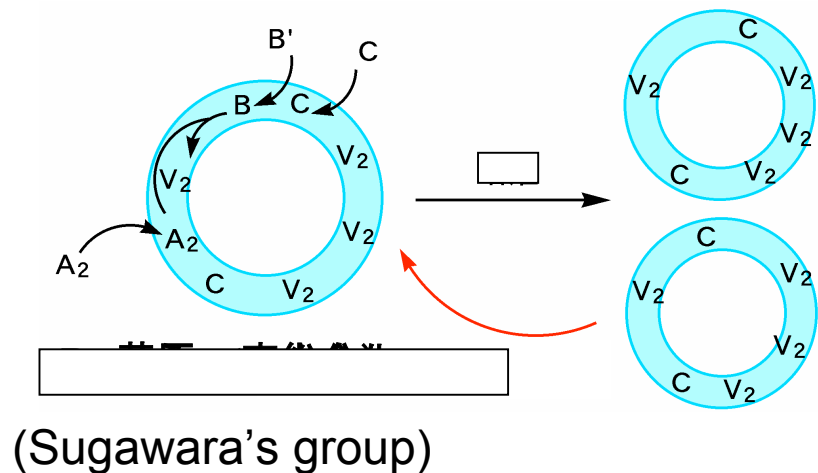
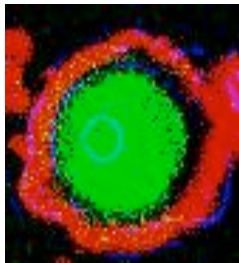


RNA polymerase



RNA polymerase gene

(Yomo's group)

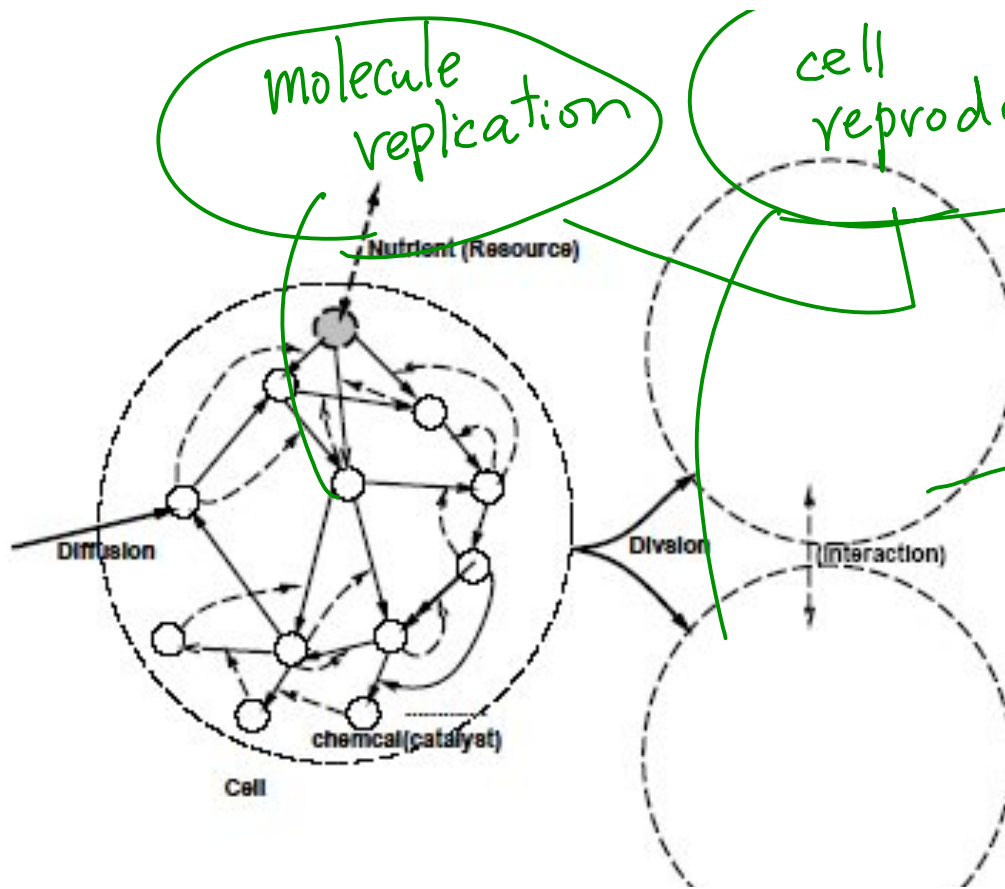


(Sugawara's group)

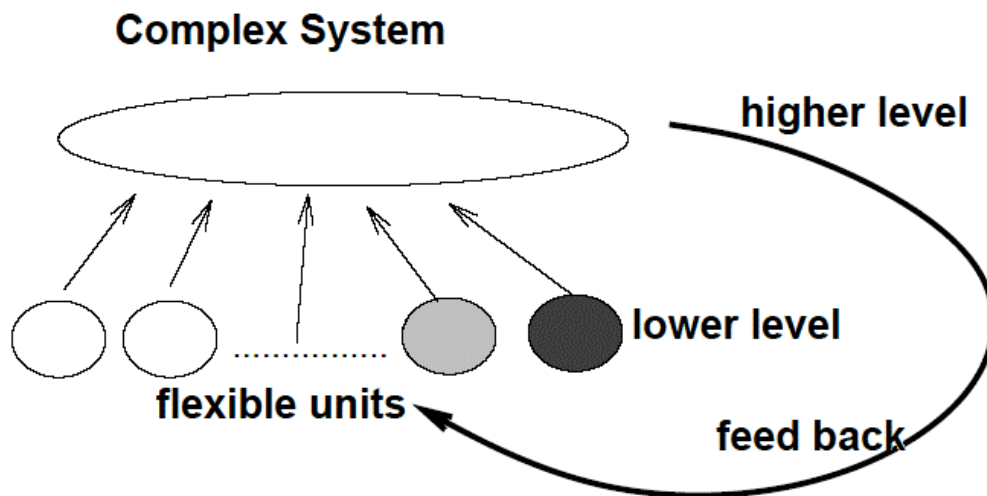
Translation in liposome

RNA replication in liposome

Continuous division of liposomes



**Consistency  
between replication of  
chemicals and  
reproduction of cell;  
keeping composition  
and activity**



**Consistency Principle for  
Biological System →  
universal characteristics**

- **Phenotypic Fluctuation →**

Relationship to Evolution?

selection is based on phenotype

(activity, size, protein abundances, fluorescence,...),

but

in standard evolutionary genetics;

gene  $a \rightarrow$  phenotype  $x$  uniquely determined

Mostly discusses the phenotype distribution  
as a result of genetic variation

——only the distribution of gene is discussed,

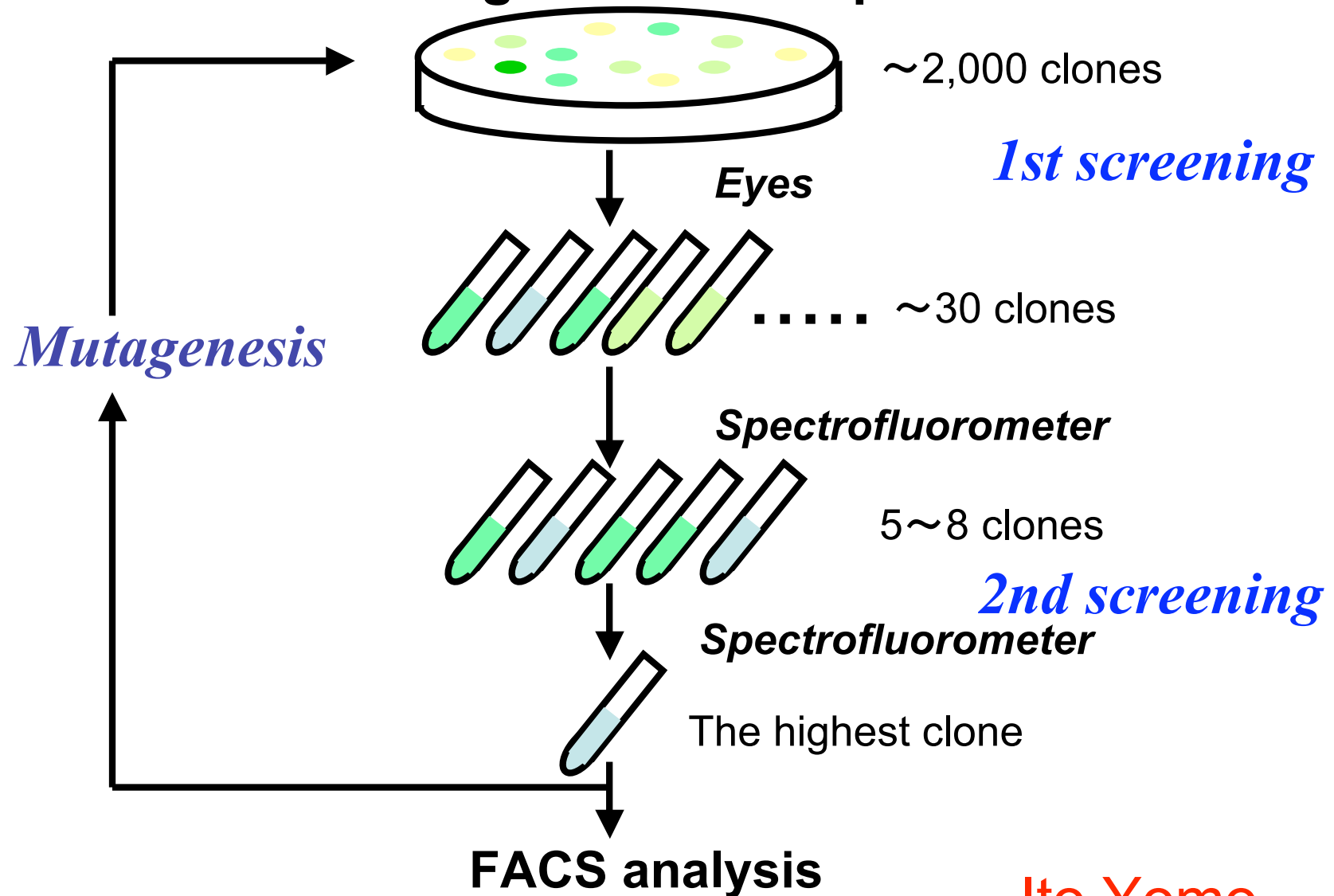
Phenotypic fluctuation of isogenic organisms

$\rightarrow P(x; a)$   $x$ —phenotype,  $a$  – gene

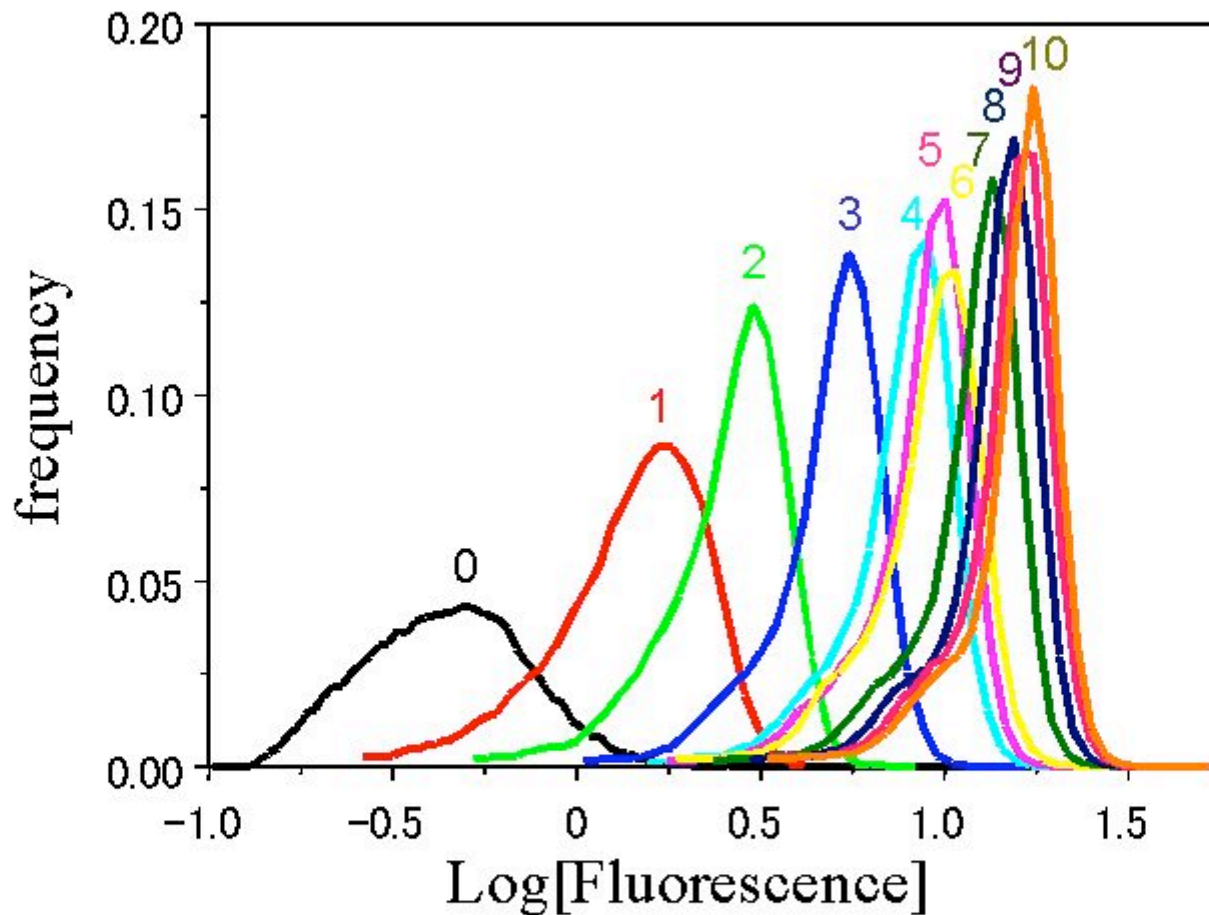
# Artificial selection experiment with bacteria

Selection to increase the fluorescence of protein in bacteria

## Schematic drawing of selection process



Ito, Yomo,...



Sato,Ito,  
Yomo,KK  
PNAS(2003)

Fluctuation --- Variance of phenotype of clone

Organisms with larger phenotypic fluctuation higher evolution speed;

- change of phenotype per generation per mutation --  
``Response against mutation+selection''

Response  $\leftrightarrow$  Fluctuation



So-called fluctuation-dissipation theorem in physics:

Force to change a variable  $x$ ;

**response ratio** = (shift of  $x$ ) / force

**fluctuation of  $x$**  (without force)

**response ratio** proportional to **fluctuation**

originated by Einstein's paper a century ago...

**Generalization::(mathematical formulation)**

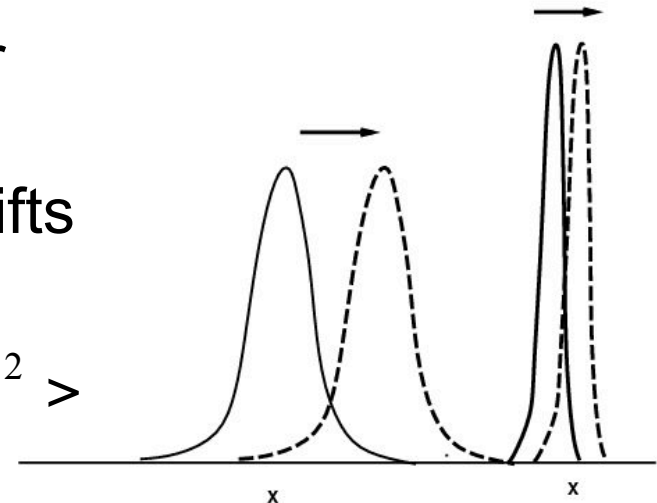
**response ratio of some variable  $x$  against the change of parameter  $a$  versus fluctuation of  $x$**

$P(x;a)$   $x$  variable,  $a$ : control parameter

change of the parameter  $a \rightarrow$

peak of  $P(x;a)$  ( i.e.,  $\langle x \rangle$  average ) shifts

$$\frac{\langle x \rangle_{a+\Delta a} - \langle x \rangle_a}{\Delta a} \propto \langle (\delta x)^2 \rangle_a = \langle (x - \langle x \rangle)^2 \rangle$$





# Fluctuation-response relationship (generalized form)

Gaussian distribution of  $x$ ; under the parameter  $a$

$$P(x; a_0) = N_0 \exp\left(-\frac{(x - X_0)^2}{2\alpha_0}\right), \quad \text{at } a=a_0$$

Change the parameter from  $a_0$  to  $a$

$$P(x : a) = N \exp\left(-\frac{(x - X_0)^2}{2\alpha(a)} + v(x, a)\right)$$

$v(a, x) = C(a - a_0)(x - X_0) + \dots$ , with  $C$  as a constant,

$$\longrightarrow P(x, a_0 + \Delta a) = N' \exp\left(-\frac{(x - X_0 - C\Delta a\alpha(a_0 + \Delta a))^2}{2\alpha(a_0 + \Delta a)}\right)$$

Hence, we get

$$\frac{\langle x \rangle_{a=a_0+\Delta a} - \langle x \rangle_{a=a_0}}{\Delta a} = C\alpha(a_0 + \Delta a),$$

Noting that  $\alpha = \langle (\delta x)^2 \rangle$

$$\frac{\langle x \rangle_{a=a_0+\Delta a} - \langle x \rangle_{a=a_0}}{\Delta a} = C \langle (\delta x)^2 \rangle,$$

Artificial selection experiment with bacteria  
for enzyme with higher catalytic activity  
for some protein with higher function

Change in gene (parameter;  $a$ )  $\Rightarrow$

“Response” ----- change of phenotype  $\langle x \rangle$

(e.g., fluorescence intensity)

per generation per (synonymous) mutation rate

Fluctuation ---- Variance of phenotype  $x$  of clone

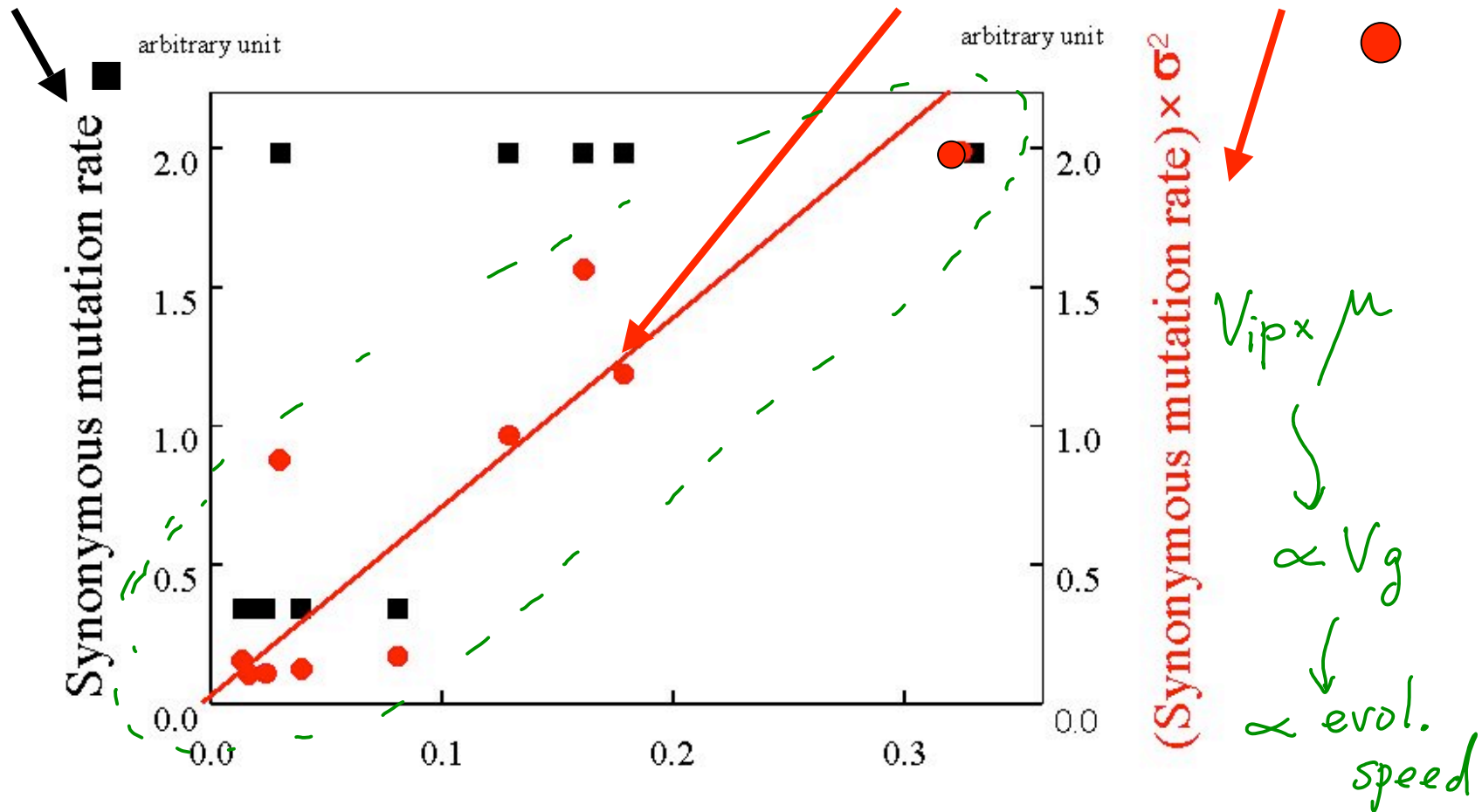
Fluctuation in the phenotype  $x$  of clone

$\Leftrightarrow$  speed of evolution to increase  $\langle x \rangle$

(proportional or correlated)

Naïve expectation:  
Just prop to mutation rate

Fluctuation-response relation  
Phenotype fluct.  $\times$  mutation rate



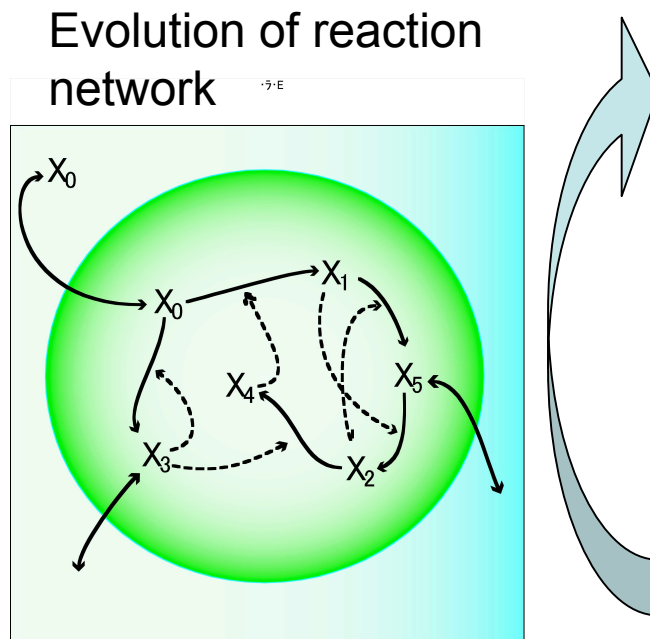
Difference of the average value

(Evolution Speed per generation)

Sato, Ito, Yomo, KK, PNAS 2003

- Confirmation by numerical evolution experiment by the reaction-net cell model

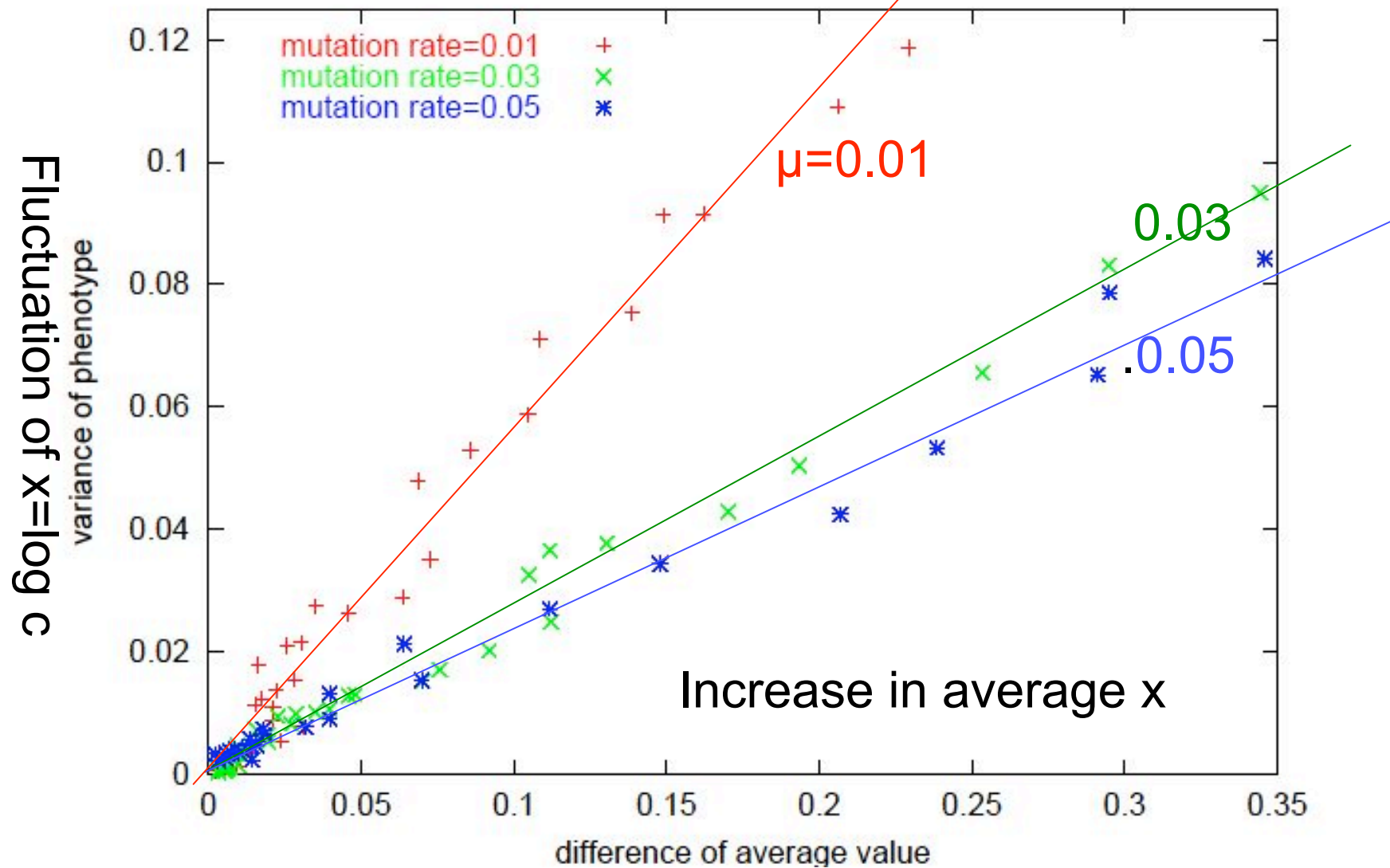
Mutate the network ('gene') with mutation rate  $\mu$ , (rewire the path of the network with the rate) and select such network having highest concentration  $c$  of a specific chemical



**phenotype  $x = \log(n_s)$**

1. Prepare initial mother cells.
2. From each parent cell, mutant cells are generated by randomly replacing reaction paths, with **mutation rate  $\mu$**
3. reaction dynamics of all mutants are simulated to determine phenotype  $x$
4. Top 5% cells with regard to phenotype  $x$  are selected as parent cells of next generation

# Confirmation of Fluctuation Dissipation Theorem by reaction-network cell model



(1) the use of  $\log(\text{fluorescence})$ , because  $\log x$  is close to Gaussian distribution in experiments

(2) New mystery? **phenotype fluctuation of clone vs evolution speed** in contrast to evolution speed  $\propto$  phenotypic fluctuation by genetic variation ( $V_g$ ): (fundamental theorem of natural selection; established)

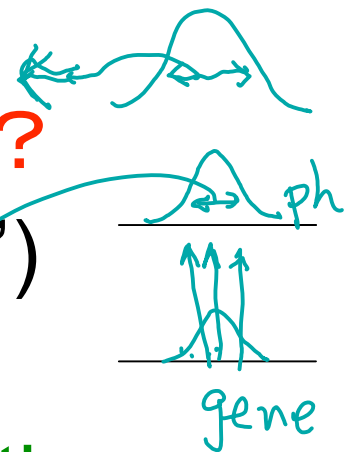
isogenic phenotypic fluct  $V_{ip}$

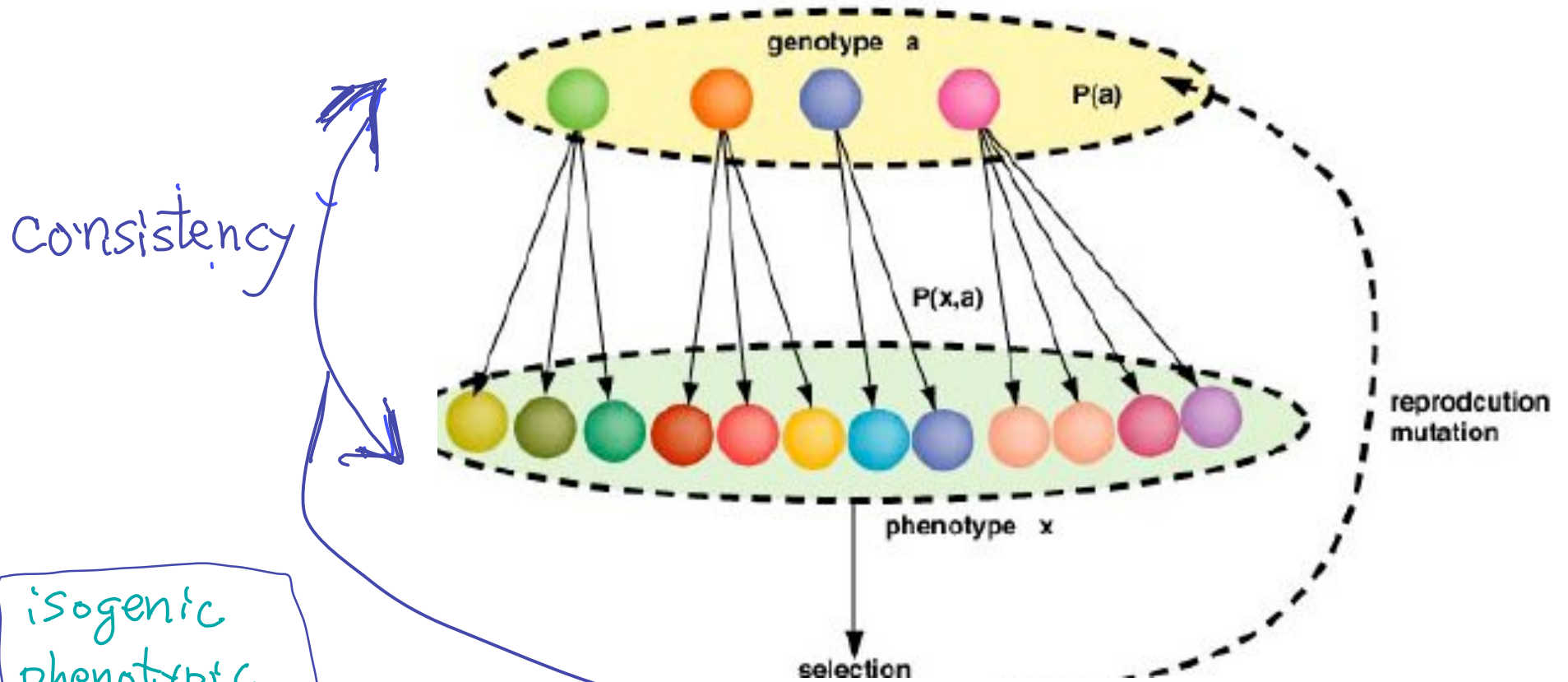
$\propto$  pheno fluct by gene variation  $V_g$ ?

(fluct by noise  $\propto$  variation in 'equation')

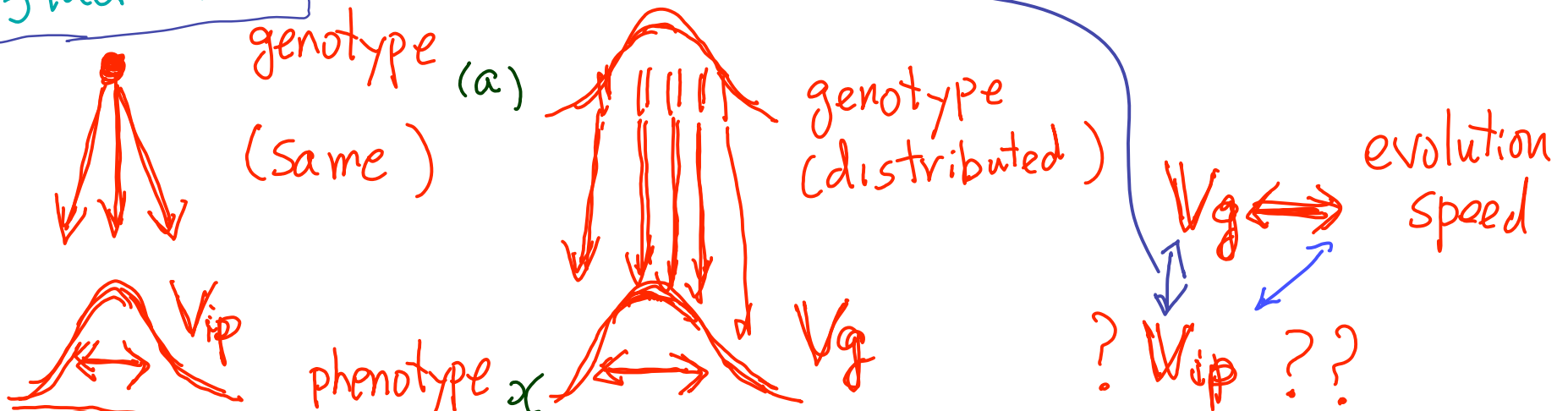
Follow the spirit of Einstein;

micro-macro consistency  $\rightarrow$  Brownian motion





isogenic  
phenotypic  
fluctuation





We can do the analysis by using Gaussian 2-body distribution function for phenotype  $x$  and gene  $a$ ; around  $a=a_0$ , and  $x=X_0$ ; with coupling between  $x$  and  $a$  (variance of  $a$  is the mutation rate  $\mu$ )

$$\underline{P(x, a)} = N \exp\left[-\frac{(x - X_0)^2}{2\alpha(a)} + C(a - a_0)(x - X_0) - \frac{1}{2\mu}(a - a_0)^2\right],$$

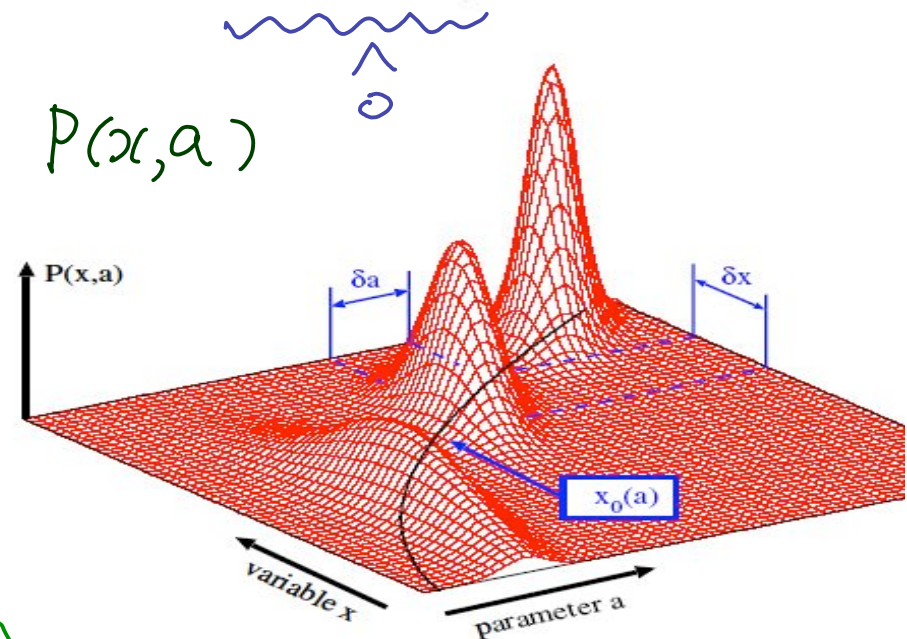
$$P(x, a) = N \exp\left[-\frac{(x - X_0 - C(a - a_0)\alpha)^2}{2\alpha(a)} + \underbrace{\left(\frac{\alpha C^2}{2} - \frac{1}{2\mu}\right)}_{\circ} (a - a_0)^2\right],$$

Stability condition

$$\frac{\alpha(a_0)C^2}{2} - \frac{1}{2\mu} \stackrel{>}{\equiv} 0, \text{ i.e.,}$$

$$\mu \leq \frac{1}{(C^2 \alpha(a_0))} \equiv \mu_c$$

For high mutation rate single-peak is not sustained (error catastrophe)





Now consider the phenotypic  
variance due to genetic variation  
Recalling the definition

$$V_g = \langle (\bar{x}_a - \bar{x}_{a_0})^2 \rangle$$

$$\bar{x}_a \equiv \int x \exp(-V(x, a)) dx = X_0 + C(a - a_0)$$

we obtain

$$V_g = \langle (\bar{x}_a - \bar{x}_{a_0})^2 \rangle = C^2 \langle (\delta a)^2 \rangle = C^2 \mu \alpha^2$$

Now the inequality  $\mu < 1/(C^2 \alpha(a_0)) \equiv \mu_c$  is rewritten as

$$V_g \leq \alpha(a_0)$$

$$V_g \leq V_{ip} \quad (1)$$

Note, in the above formulation,  $\langle (\delta a)^2 \rangle = \mu$  and  $V_g \propto \mu$ . Recalling that  $V_g$  at  $\mu_c$  equals  $V_{ip}$ , we get

$$V_g = \frac{\mu}{\mu_c} V_{ip}$$

$$\rightarrow V_g \propto V_{ip} \quad (3)$$

**Error Catastrophe**  
**As  $V_g \geq V_{ip}$  (2)**

Fisher's theorem : evolution speed  $\propto V_g$ ,  
EFR : evolution speed  $\propto \mu V_{ip} \leftarrow$  Experiment

- Three general laws

- (i)  $V_{ip} \geq V_g$

- (ii) error catastrophe at  $V_{ip} \sim V_g$

- (where the evolution does not progress)

- (iii)  $V_{ip} \propto \mu V_g$  ( $\propto$  evolution speed)

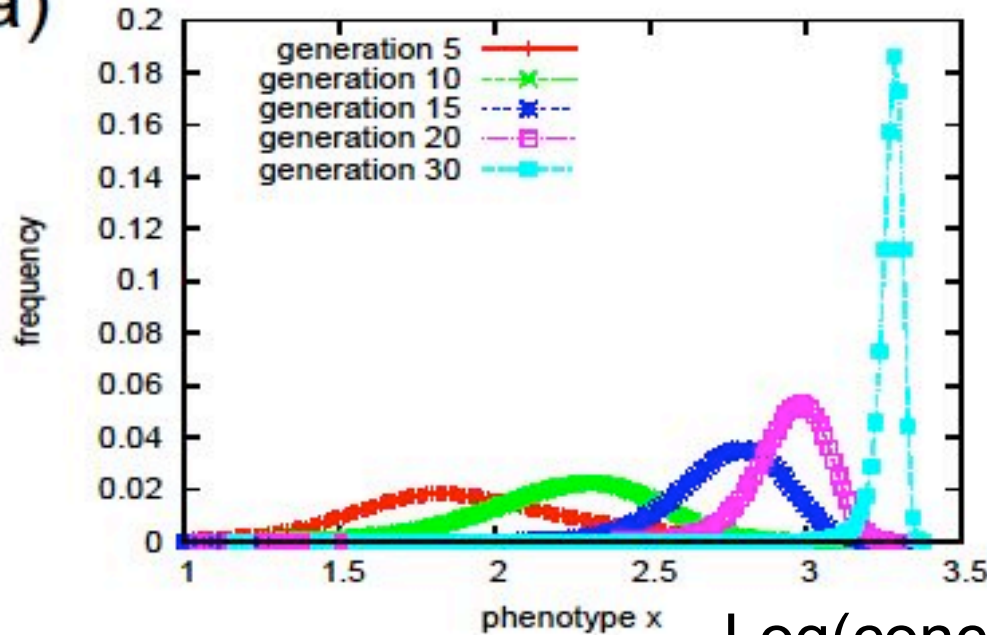
Relation (iii) seems to be fine ... but ...

relations (i) (ii) are rather surprising

----need confirmation

→ previous reaction-net cell model

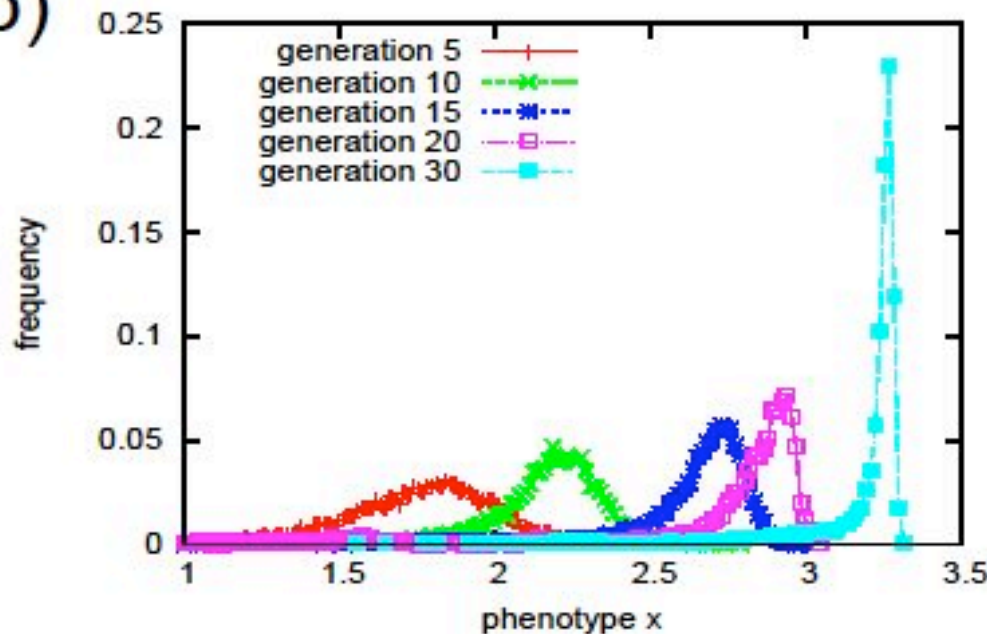
(a)



Change in distribution  
through evolution

Distribution of phenotype  
 $x$  of a clone  
 $\rightarrow V_p$

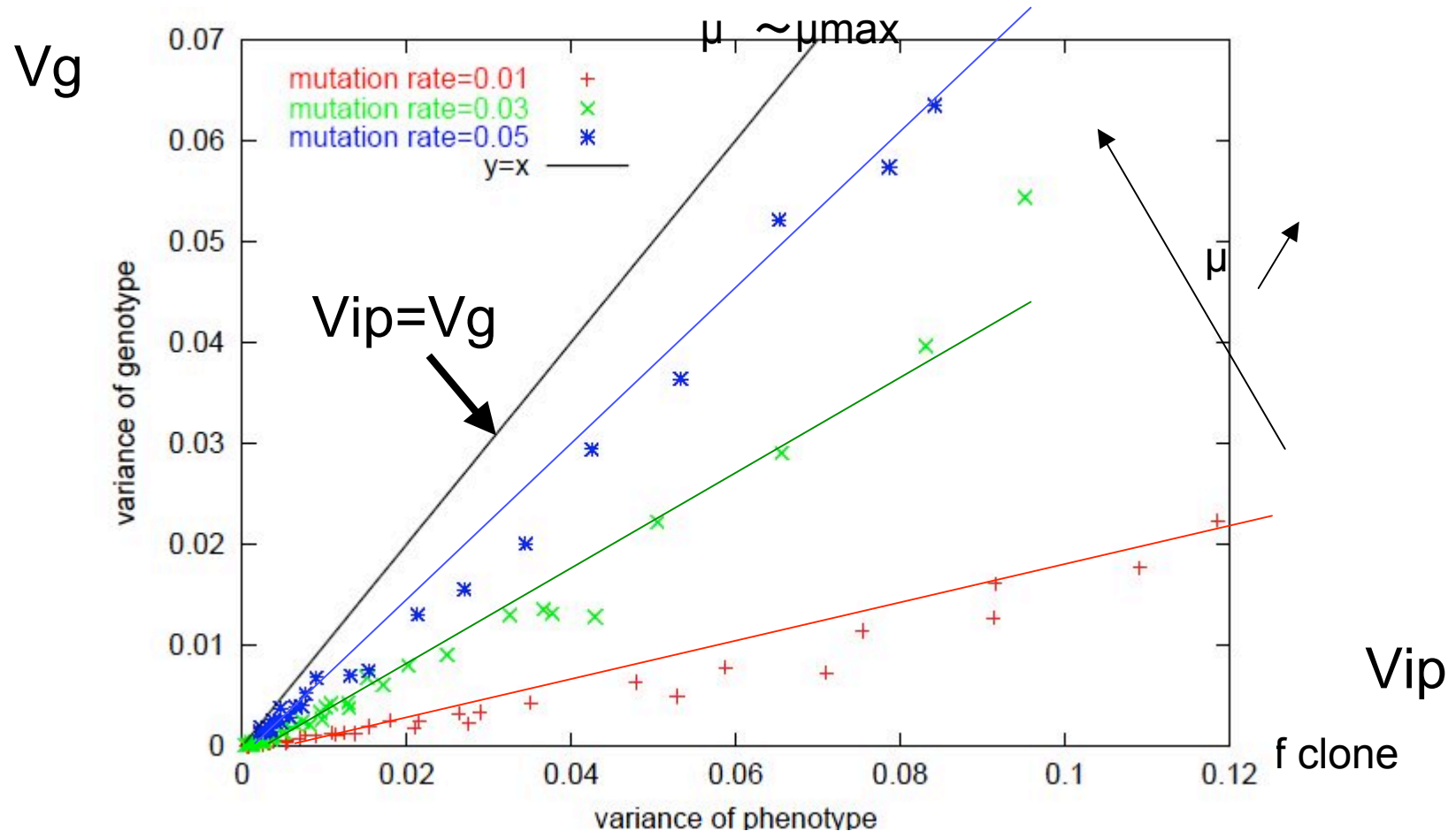
(b)



Distribution of phenotype  
 $x$  over mutants (genetic  
variation)  
 $\rightarrow V_g$

Phenotype fluct. ( $V_p$ ) vs Gene Fluct. ( $V_g$ ) in the evolution of toy cell model

$V_p$ : fluct. for given network,  $V_g$ : fluct. by network variation



variance of  $\log(x)$ ,  
 $x$  is the concentration of the molecule

Beyond Darwin with the spirit of Einstein!

As  $\mu$  (mutation rate) increases to  $\mu_{\max}$ ,

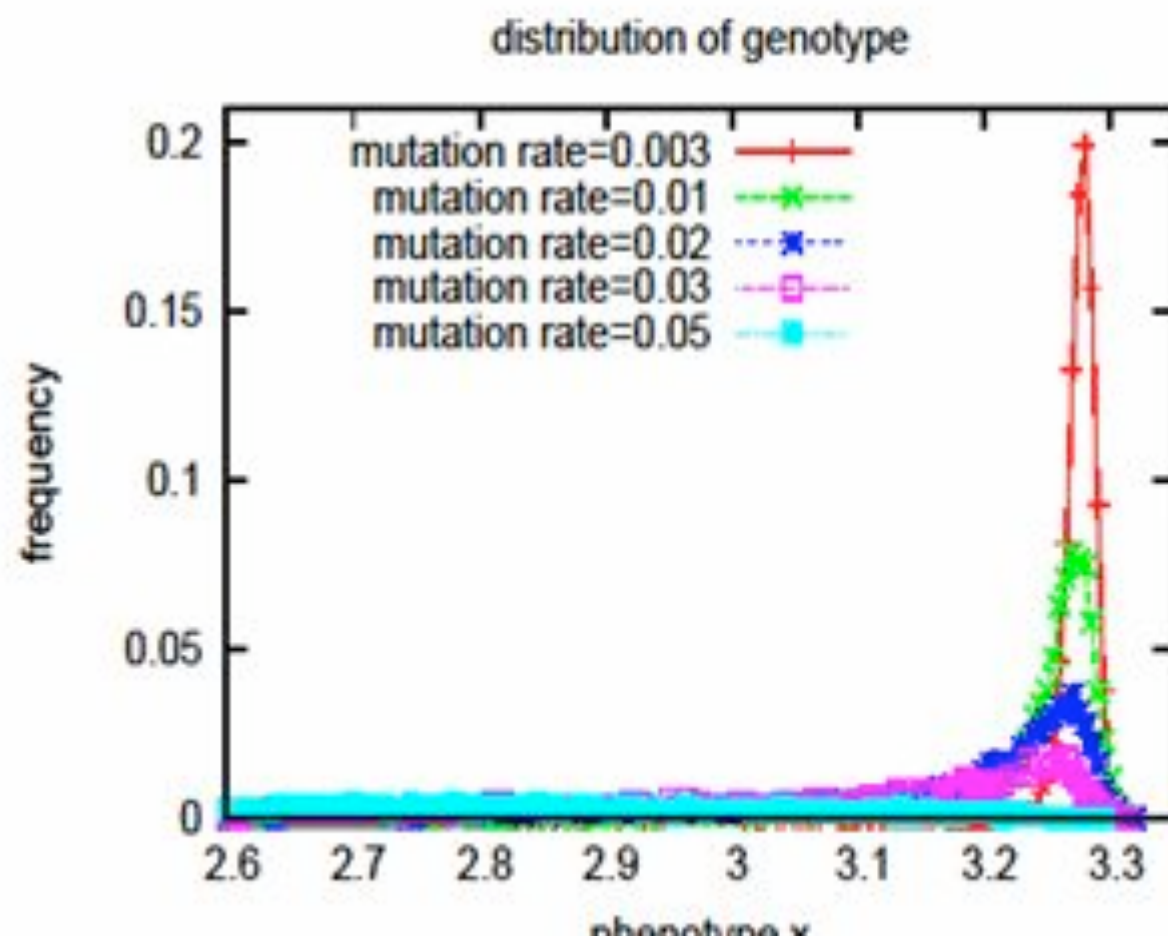
(1) the distribution collapses (error catastrophe)

(2) evolution no longer progresses beyond  $\mu_{\max}$   
evolution speed is maximal at  $\mu \sim \mu_{\max}$

(3)  $V_g$  approaches  $V_p$

As  $\mu$  is increased,  
The distribution  
'collapses'

Error catastrophe



- **Still,,??? to the theory**
- $P(x,a)$  rather than conditional probability (TRICK)

“Genetic-Phenotypic correspondence”

what phenotype can vary  $\leftrightarrow$

what gene can change

fluctuation of variable (micro) vs

variation of equation (genetic evolution)

(cf Waddington’s genetic assimilation)

**Q: Why error catastrophe when  $V_g > V_{ip}$ ?**

Robust evolution is possible only under noise

-**counterintuitive ;it says phenotype noise is important**

→ gene-net model

## A simple model for Geno-Pheno relationship;

Model: Gene-net (dynamics of stochastic gene expression)  $\rightarrow$  on/off state

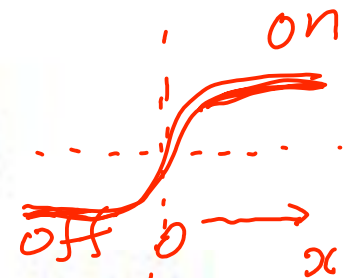
$X_i$  – expression of gene  $i$  : + (on) – (off)

$$dx_i/dt = \tanh\left[\beta \sum_{j>k}^M J_{ij}x_j\right] - x_i + \sigma\eta(t),$$

$$J_{ij} = -1, 1, 0,$$

$$\langle \eta(t)\eta(t') \rangle = \delta(t-t').$$

Gaussian white



$M$ ; total number of genes,  $k$ : output genes

Noise strength  $\sigma$



- Task

Starting from  $-1, -1, -1, \dots, -1$  (all off)

$x_i$   $i=1, 2, \dots, k$  are  $+1$  (on) (Target Gene Pattern)

**Fitness**  $F = -(\text{Average number of off } x_i)$

$\overline{\phantom{x}}$  is temporal average between  $t = T_{ini}$  and  $t = T_f$

## Genetic Algorithm

Select networks with higher  $\langle F \rangle$

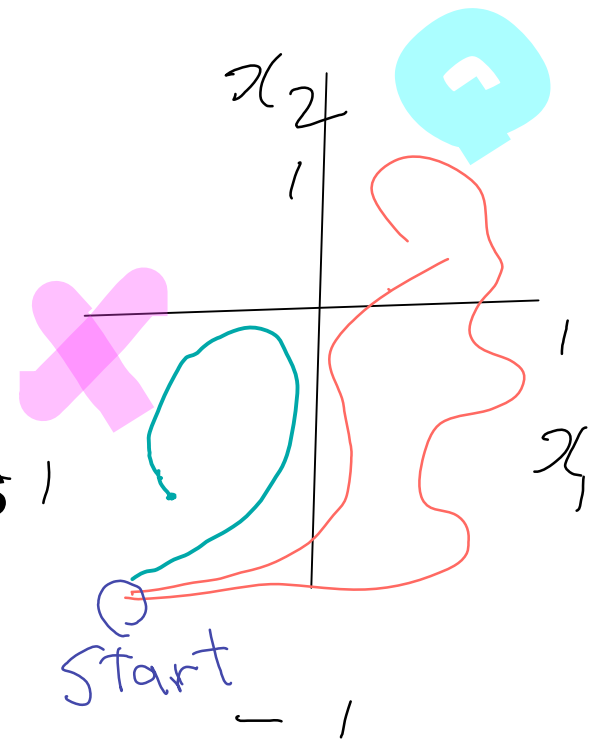
top-- $\langle F \rangle = 0$

Choose top  $n$  networks

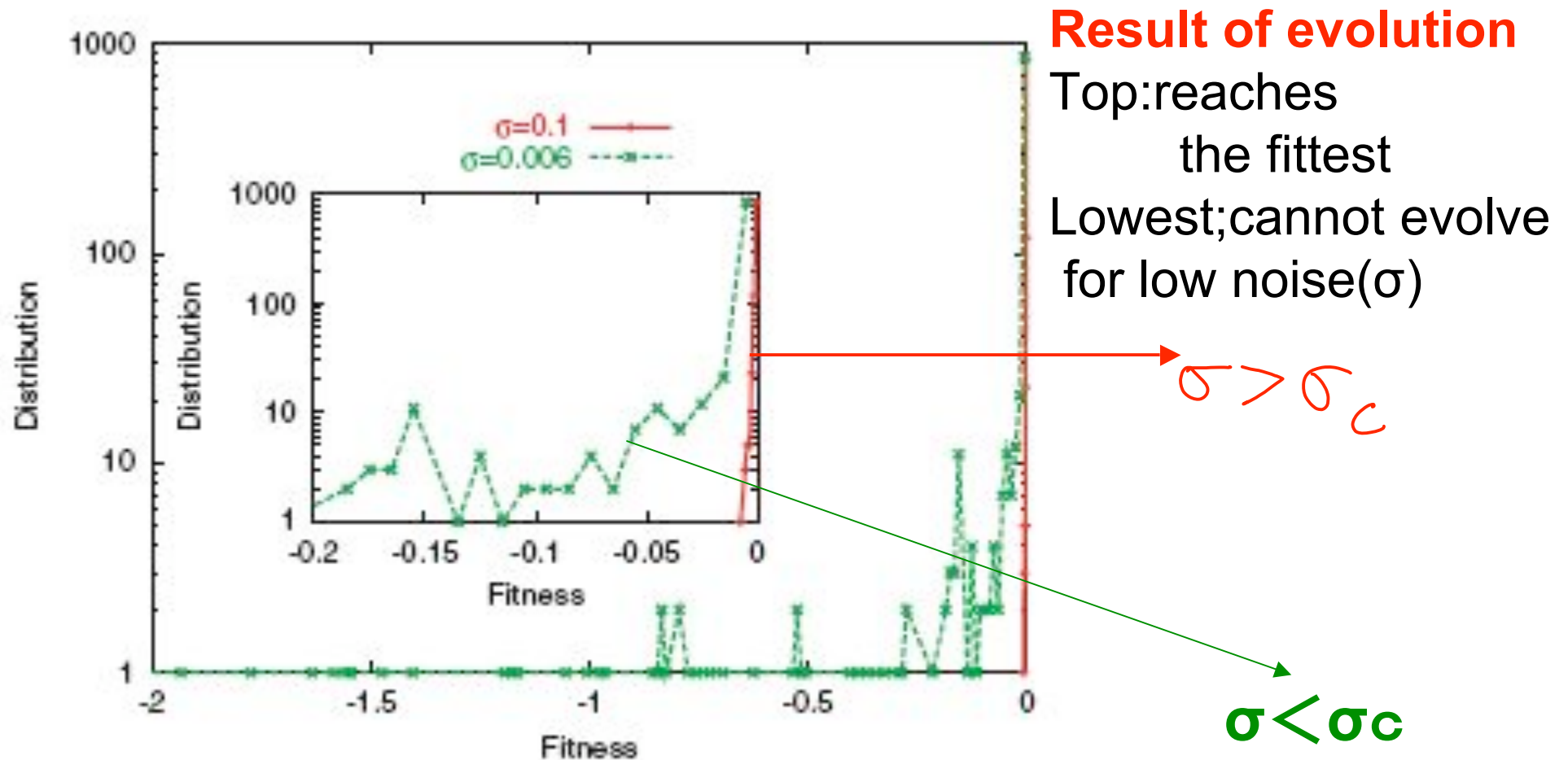
among total  $N$ , and mutate

with rate  $\mu$  to produce  $N$  networks

( $\mu$ : fixed mutation rate)





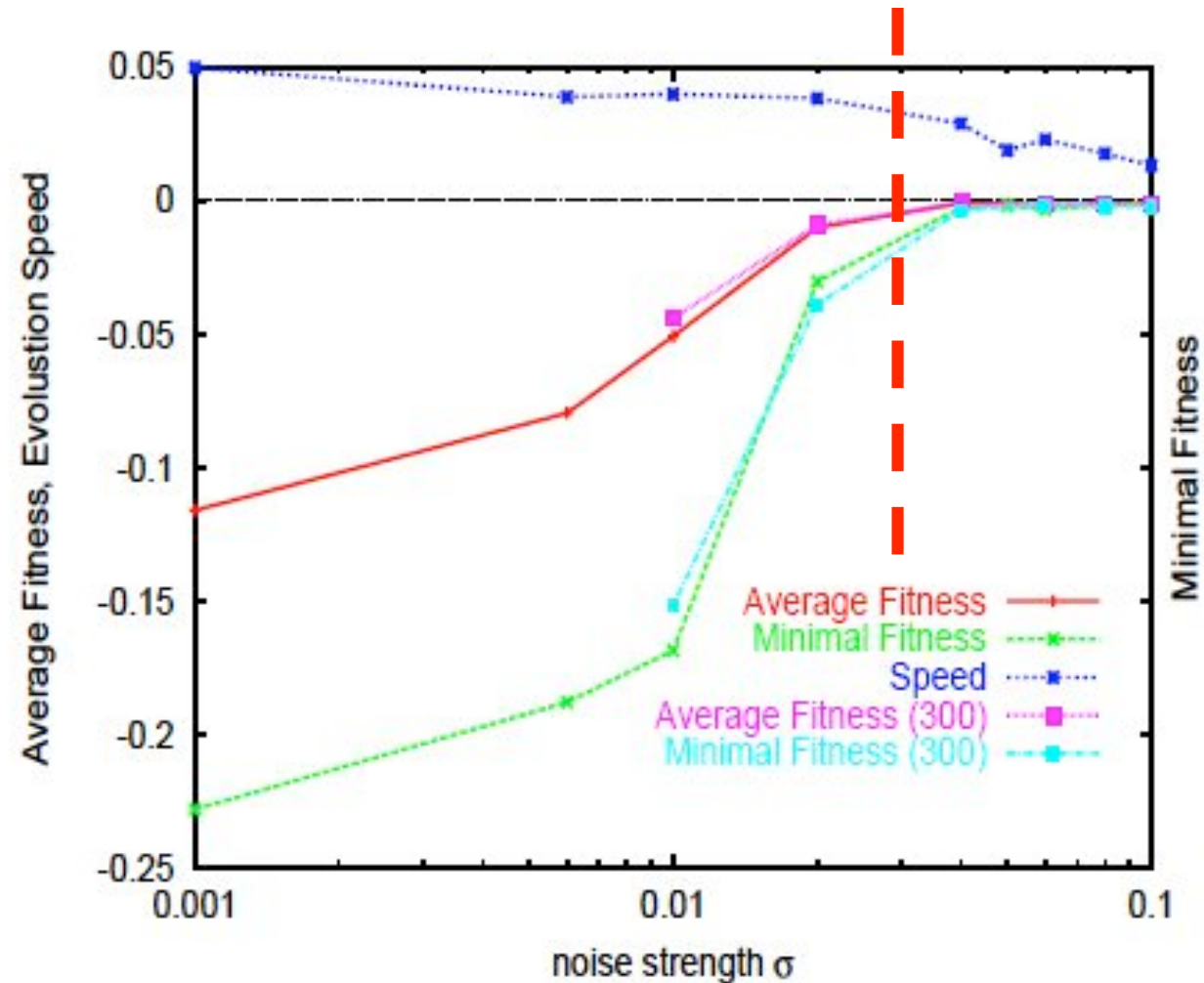


## Fitness Distribution

$\sigma < \sigma_c$  --low fitness mutants distributed

$\sigma > \sigma_c$  — eliminated

through evolution



**Existence of critical noise level  $\sigma_c$   
below which low-fitness mutants accumulate  
(error catastrophe)**

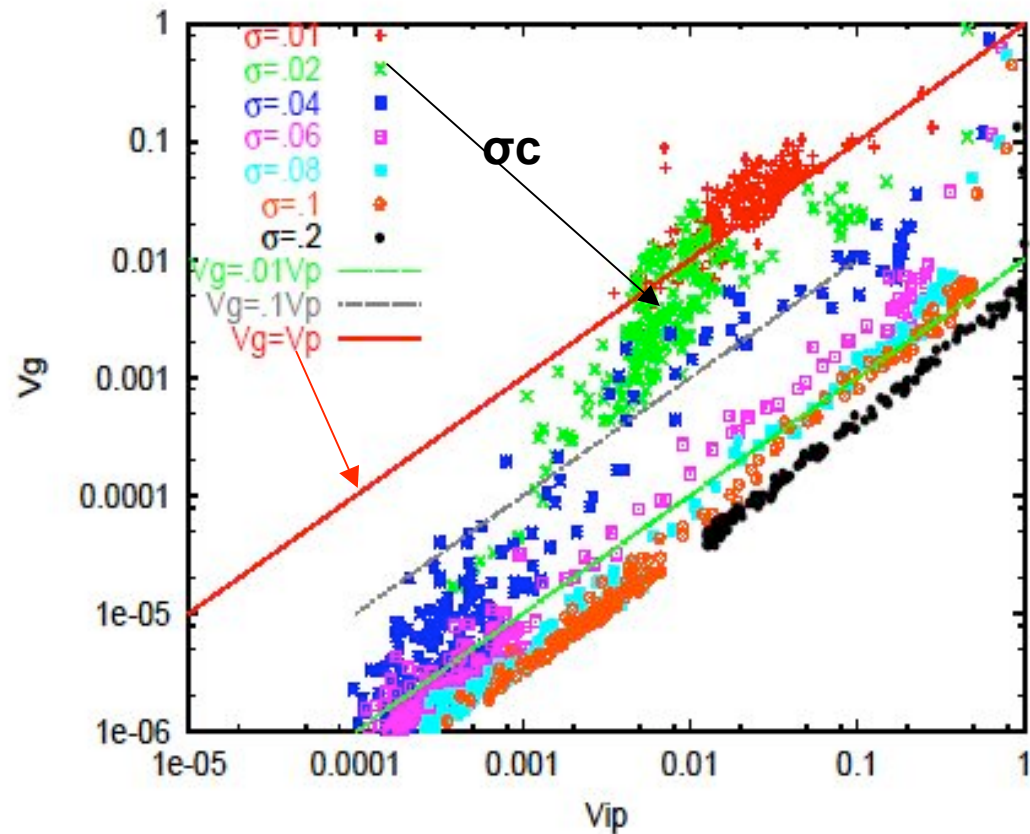
(1)  $V_{ip} \geq V_g$  for  $\sigma \geq \sigma_c$

(2)  $V_g \rightarrow V_{ip}$   
as  $\sigma \rightarrow \sigma_c$

(3) evolution progresses  
only for  $V_{ip} \geq V_g$

(4)  $V_{ip} \propto V_g$   
through evolution course (✓)

Theory confirmed

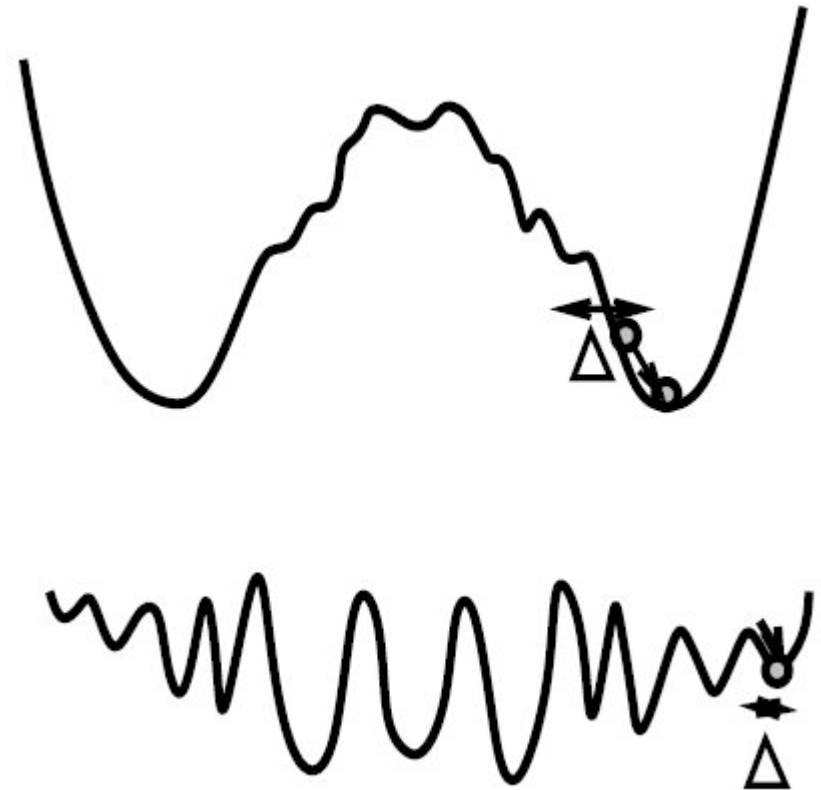
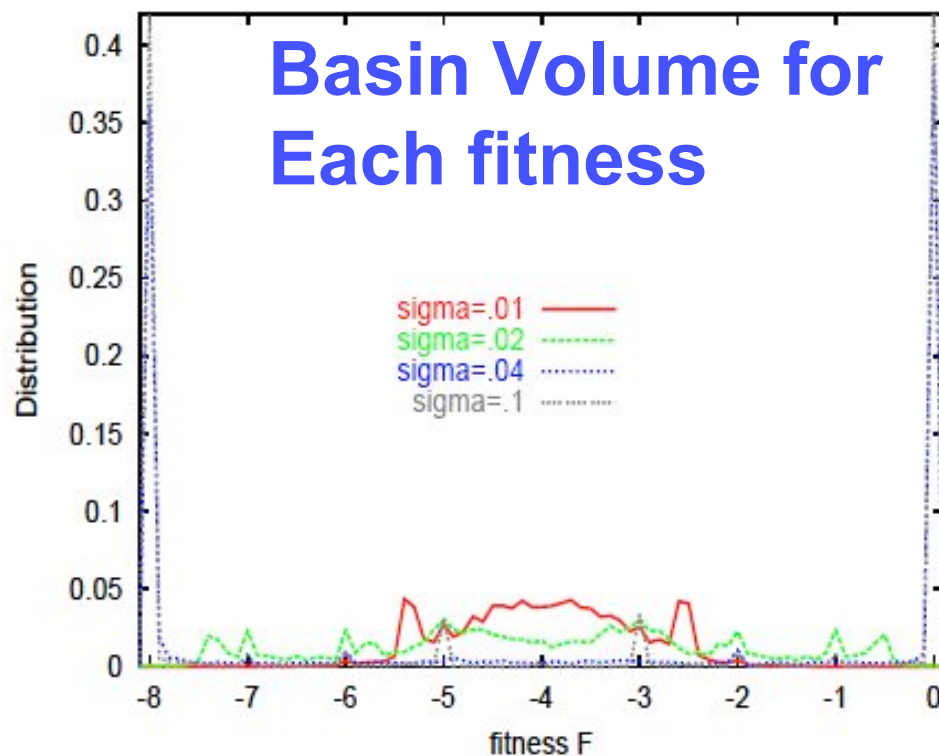


KK, PLoSOne, 2007

**Why?; difference in basin structure**

$\sigma > \sigma_c \rightarrow$  large basin for target attractor  
(robust,  $\Delta$ (distance to basin boundary)  $\uparrow$ )

$\sigma < \sigma_c \rightarrow$  only tiny basin around target orbit  
 $\Delta$  remains small



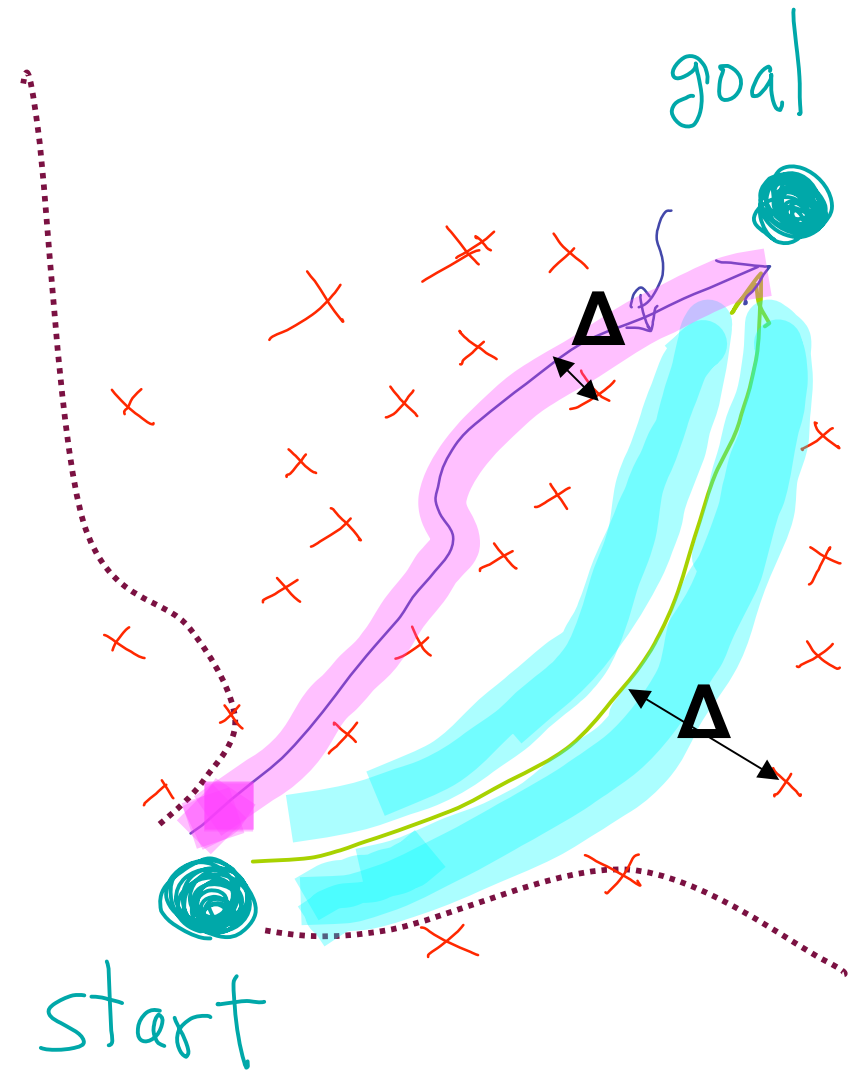
**$\rightarrow$ Global constraint to potential landscape(funnel?)**



why threshold?

choose paths to avoid turning  
pts within  $\sigma$  (noise)

Mutation  $\rightarrow$  touches turning  
points within range of  $\mu$

small  $\sigma \rightarrow$   
an orbit with small  $\Delta$   
can reach the target



	$\Delta$	low ( $\leq \sigma_c$ )
	$\Delta$	high ( $\geq \sigma_c$ )

Deviation of basin  
boundary (turning points)  
by Noise  $\rightarrow \delta p$   
by Mutation  $\rightarrow \delta g$

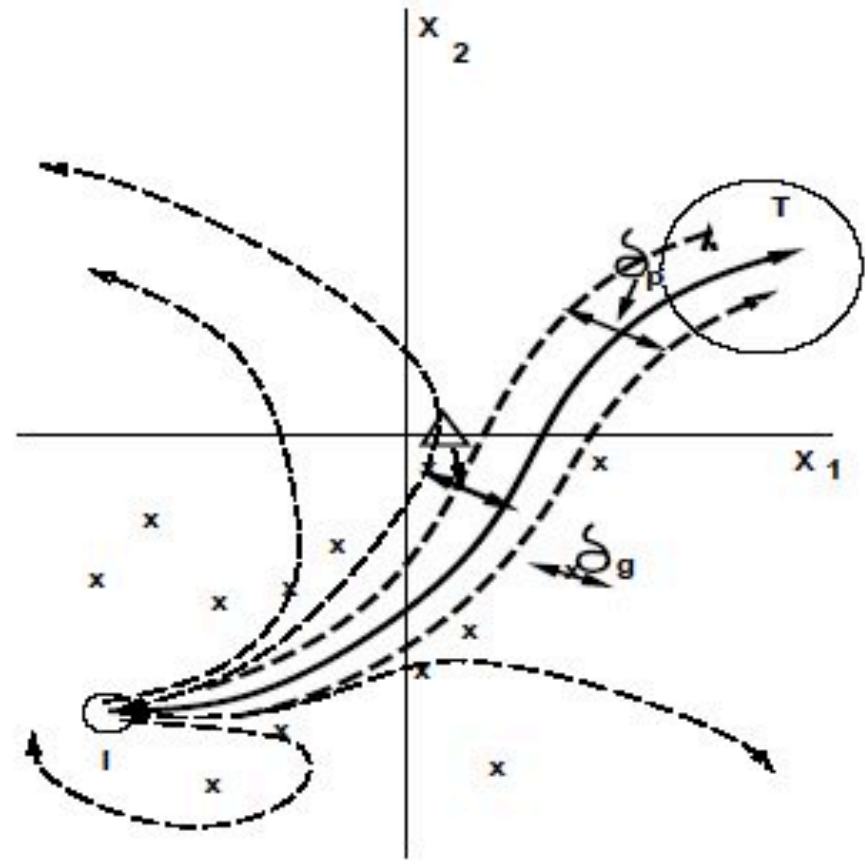
$$V_g \sim (\delta g / \Delta)^2$$

$$V_{ip} \sim (\delta p / \Delta)^2$$

$\Delta$  increases  
 $\rightarrow$  robustness  
increases

if  $\delta g > \delta p$ ,  
mutation destroys  
the history

$\rightarrow V_{ip} > V_g$  necessary  
for evolution of robustness



$\Delta \sim$  distance to turning points  
(basin boundary)

- Generality of our result; For a system satisfying:
  - (1) fitness is determined after developmental dynamics
  - (2) developmental dynamics is complex  
(catastrophic pts leading to error are distributed)
  - (3) effective equivalence between mutations and noise with regards to the consequence to fitness  
(→ genetic assimilation by Waddington)

# Discussion: Evolution of Robustness

- Robustness ----- Insensitivity of Fitness (Phenotype) to system's change
  - ← against noise during 'developmental process
  - ← against parameter change by mutation
- Developmental Robustness to noise ----  $V_{ip}$
- Robustness to mutation in evolution ----  $V_g$

When  $V_{ip} > V_g$ , both decrease, i.e., robustness ↗

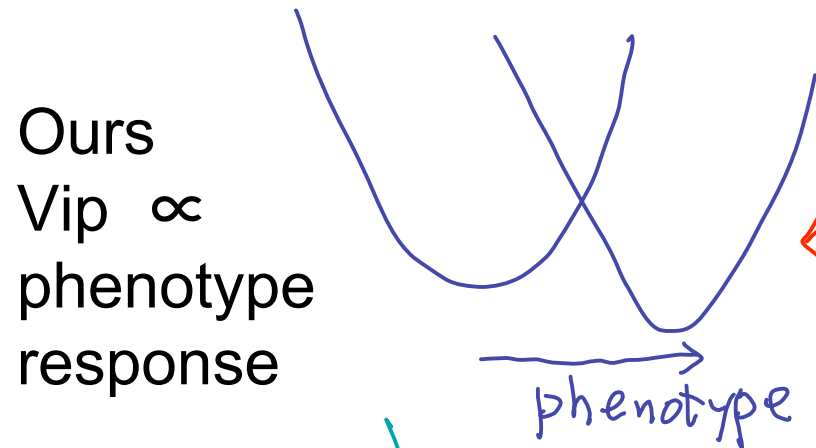
Noise is necessary for evolution of robustness

$V_{ip} \propto V_g \rightarrow$  Developmental robustness and genetic (evolutionary) robustness are linked  
(WADDINGTON)



# LeChatlier-Braun principle, Waddington's 1957?, and Vg-Vip relationship

External change → Response to suppress the influence



$V_g \propto$   
genetic response

← stability condition from  
thermodynamic potential

phenotype change by  
environmental change  
(without genetic)

↓ evolution  
(genetic change)

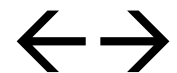
after environmental change -  
is cut off, change remains  
(buffered to gene)

- Nature vs Nurture?
- Standard population genetics:  
non-genetic variations are regarded to be due to environmental variation instead of fluctuation
- The ratio of genetic variation to total variation is called “heritability”. This value, for most cases is less than .5 (cf: data in Drosophilla 0.2-0.5)
- Our argument shows heritability  $< 1/2$ , as  $\text{heritability} = V_g / (V_{ip} + V_g)$  (if  $V_{ip}$ ,  $V_g$  are added independently) by regarding  $V_{ip}$  as origin of non-genetic variation  
→ (?Nature < Nurture?) for phenotype relevant to fitness

Through directed evolution; fluctuations decrease

(\*\*Model, experiments, theory, i.e., increase of robustness through evolution.)

Then, evolution slows down..



How Evolution continues?

Why Large Fluctuations exist?

?? Is there regain of fluctuations????

- Observed: Appearance of mutants with large fluctuations (due to different source) at further evolution. (← interaction with other genes?)
- → Restoration of Plasticity

# Spontaneous Adaptation

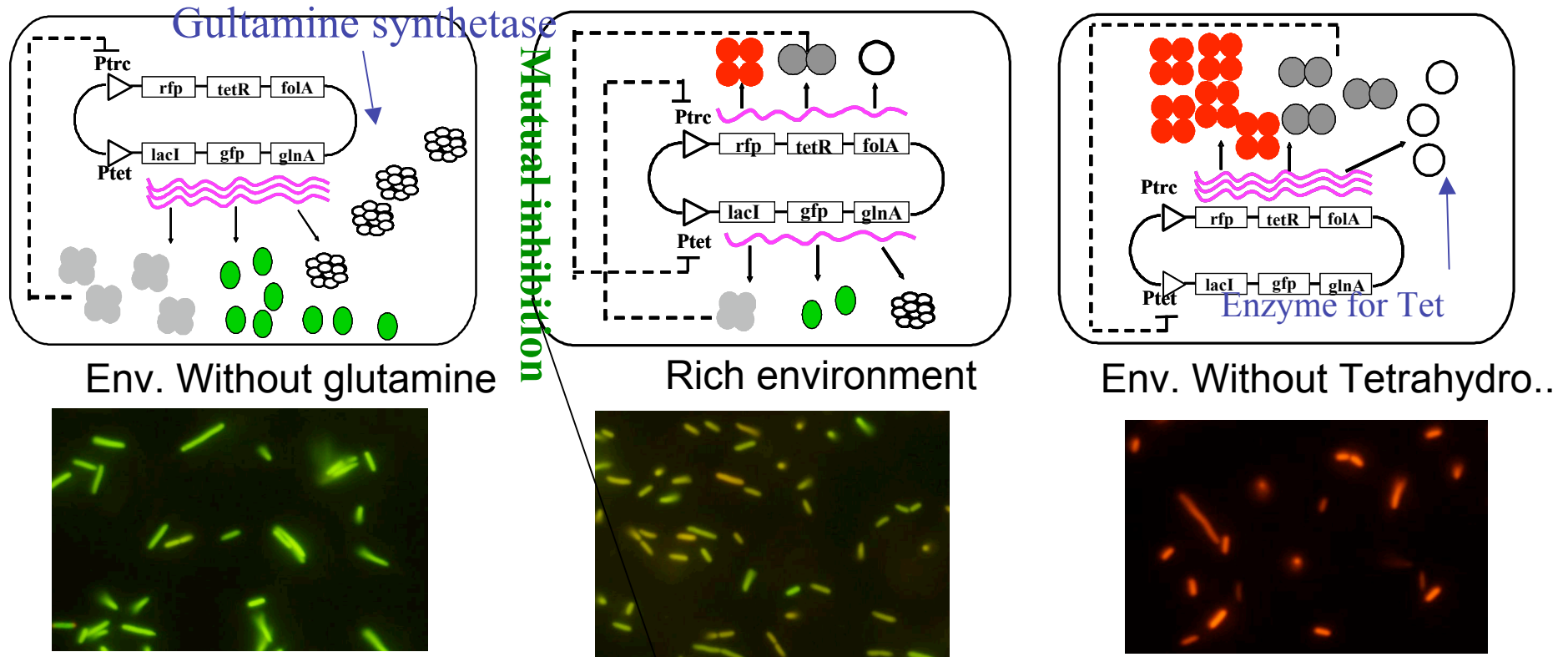
- For all possible changes in environment, signal transduction network is already provided?
- Or, is there any general (primitive) mechanism to make spontaneous adaptation?
- → Constructive Experiment with artificial Gene and theory assuming only growth condition and stochasticity
- From consistency between cellular growth and stochastic gene expression dynamics, adaptive attractors are generally selected (theory)

# (ex) Adaptive response without signal transduction

Embedded gene network

Unexpected; beyond designed  
Selection of preferable state

Phenomenological theory of attractor selection



fluctuation

Metabolic activity

Theory of attractor selection by  
activity and noise

Kashiwagi, Yomo

- Growth-Induced-Attractor-Selection (Furusawa kk)
- Basic Logic

$$dx_i/dt = f(x_i) - S(\{x_j\})x_i + \eta(t)$$

$f \rightarrow$  Synthesis     $S \rightarrow$  dilution due to cell growth

$\eta \rightarrow$  noise

**Active state** : both  $f$  and  $S$  are large

deterministic part  $\gg$  noise

**Poor state** : both  $f$  and  $S$  are small

deterministic part  $\sim$  noise

Switch from Poor state to Active state by noise

Selection before reproduction

General logic in a system with growth and fluctuation

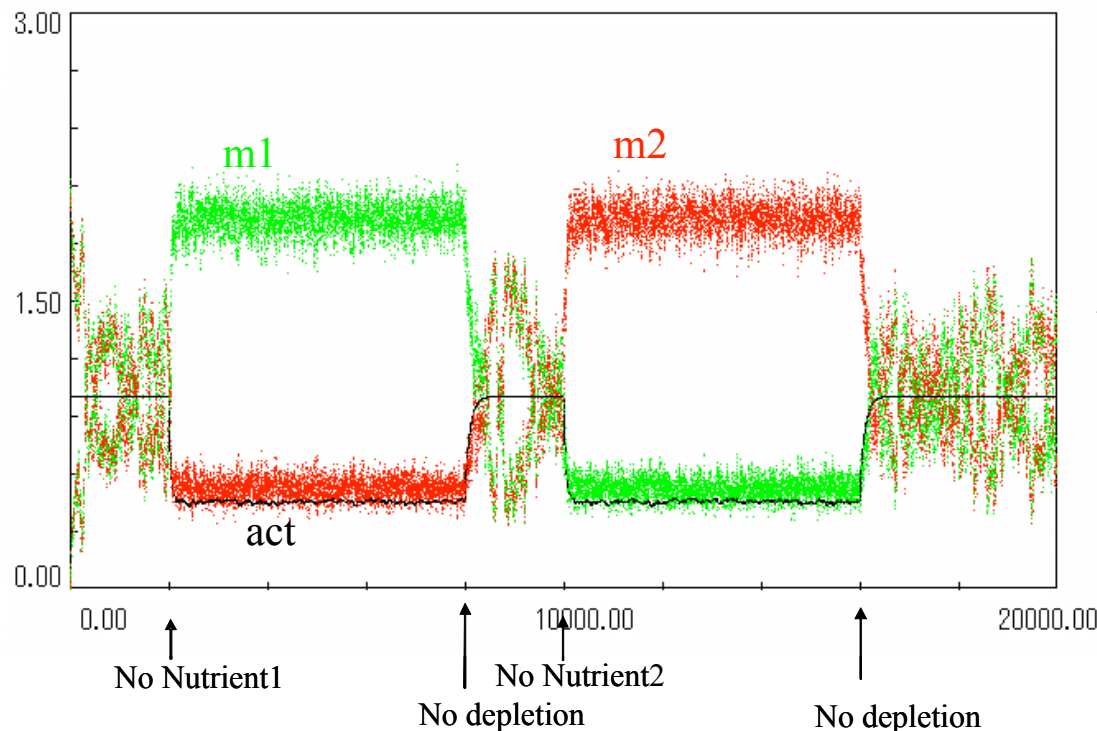
# The mechanism for adaptive response by attractor selection

$$\frac{d}{dt}m1 = \frac{\text{syn}(act)}{1+m2^2} - \text{deg}(act) \times m1 + \eta_1$$

$$\frac{d}{dt}m2 = \frac{\text{syn}(act)}{1+m1^2} - \text{deg}(act) \times m2 + \eta_2$$

$$\text{syn}(act) = \frac{6act}{2+act}; \text{deg}(act) = act;$$

$$\frac{d}{dt}act = \frac{\text{pro}}{\left(\left(\frac{\text{Nut\_thread}_1}{m1 + \text{Nutrient1}}\right)^{n_1} + 1\right) \times \left(\left(\frac{\text{Nut\_thread}_2}{m2 + \text{Nutrient2}}\right)^{n_2} + 1\right)} - \text{cons} \times act$$



Adaptive Response of the genetic network to a environmental change





# Topic4; Cell differentiation: Isologous Diversification: (KK,Yomo1997)

$$\frac{dx^m}{dt} = f_m(x^1, x^2, \dots, x^k)$$

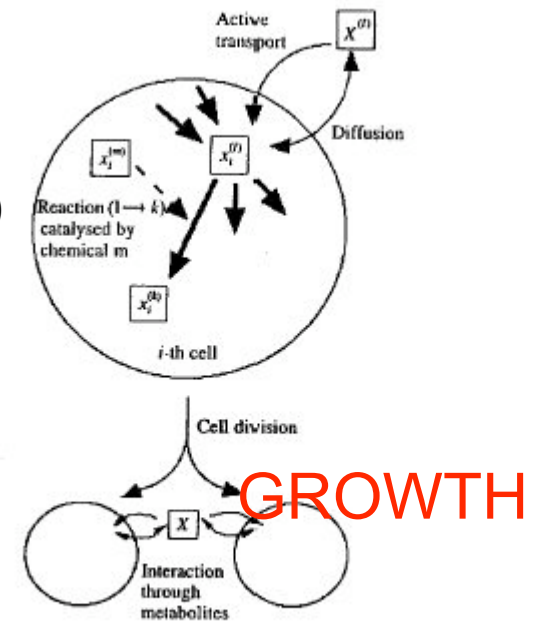


FIG. 1. Schematic representation of our model. See the appendix for the specific equation of each process.

Reproduction of a cell  
vs growth as a multicellular organism  
→ development

Internal chemical reaction dynamics  
and interaction and cell division

Assuming oscillatory reaction dynamics.,

+GROWTH (→ change phase space dimension)

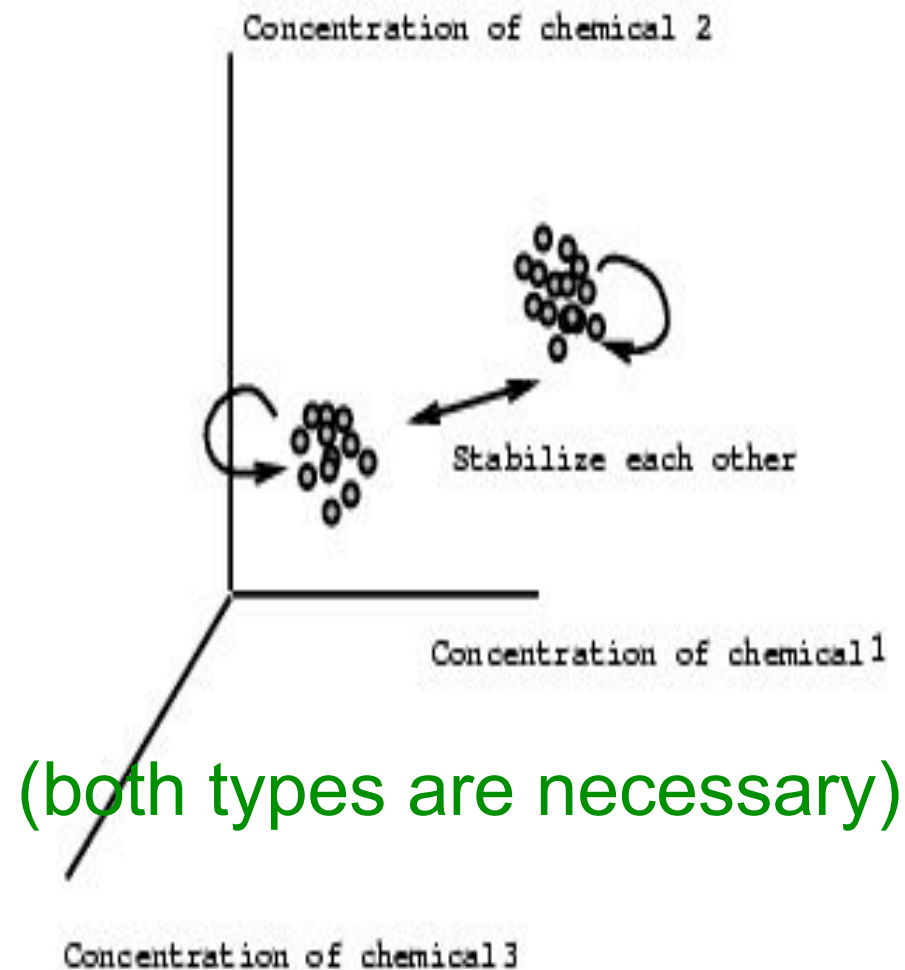
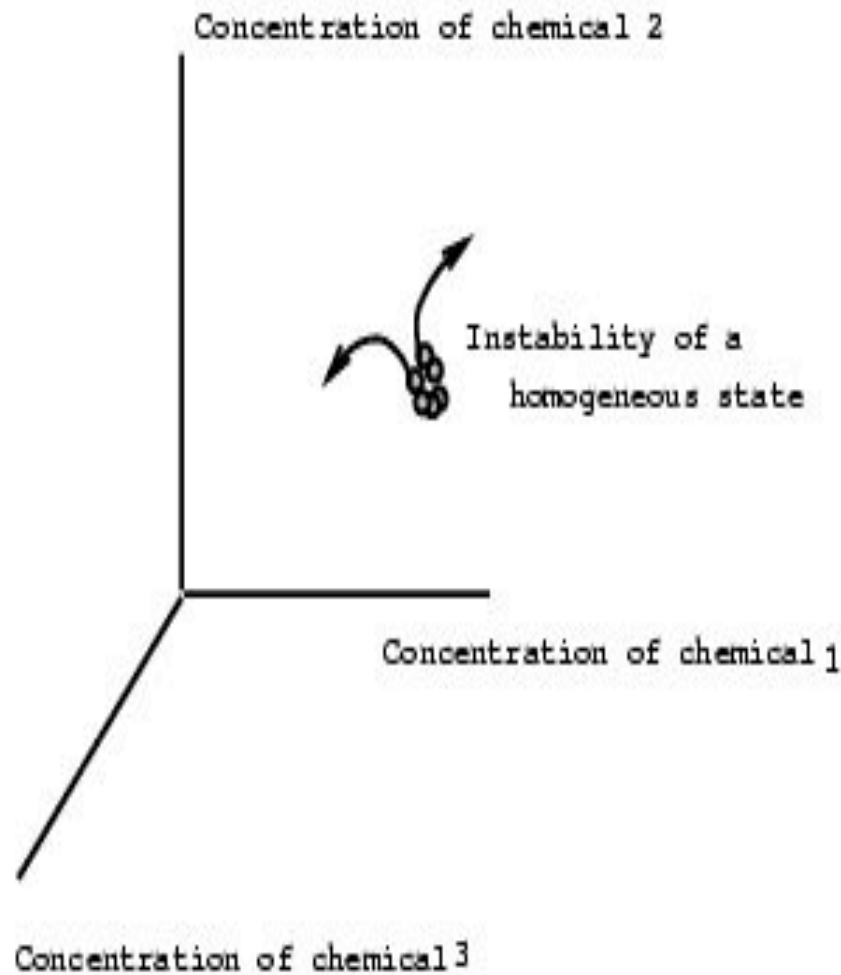
Cell number increases → interaction change

→ Bifurcation of intra-cellular dynamics → differentiation

Stem Cell (chaotic dynamics) → stochastic

differentiation with spontaneous regulation of probability  
to keep the consistency between cell and population

→ With the increase of the number

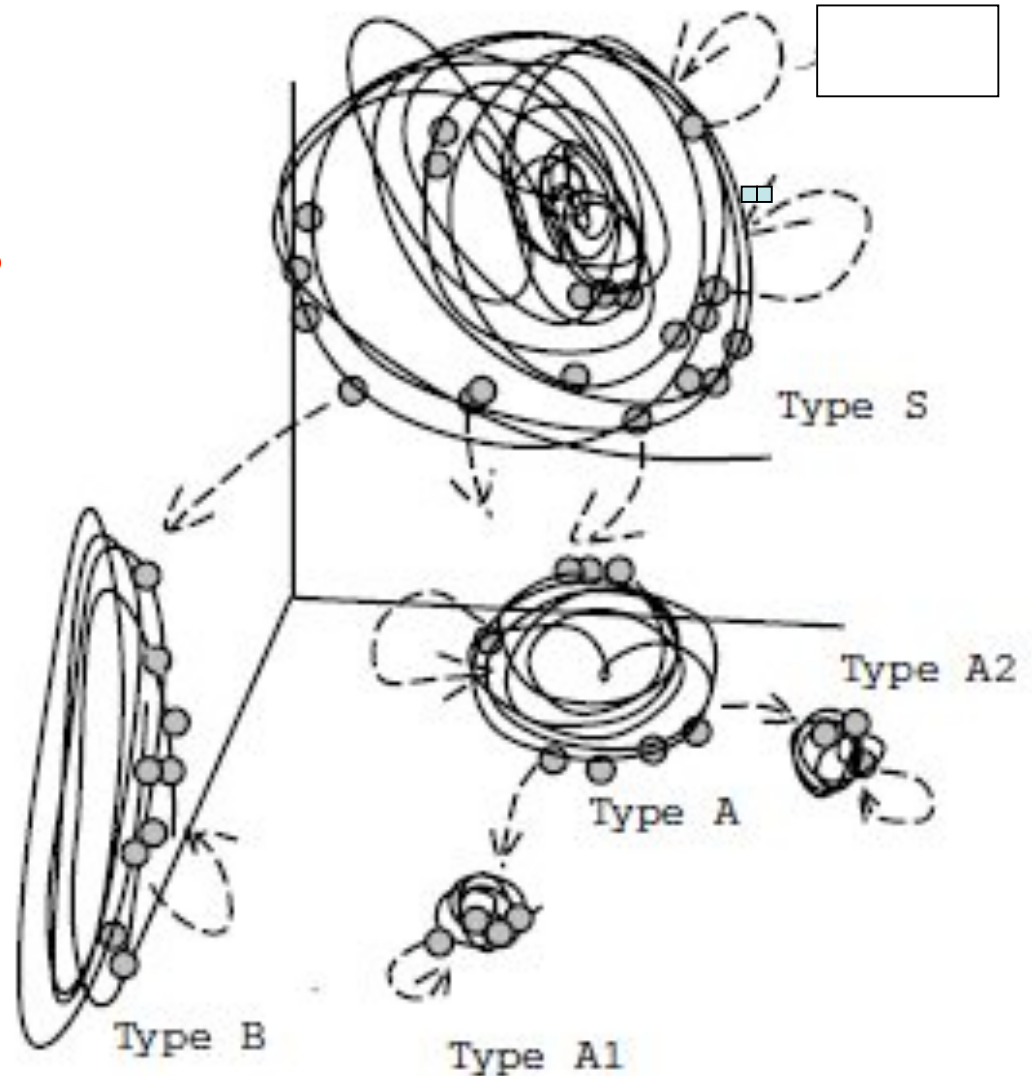
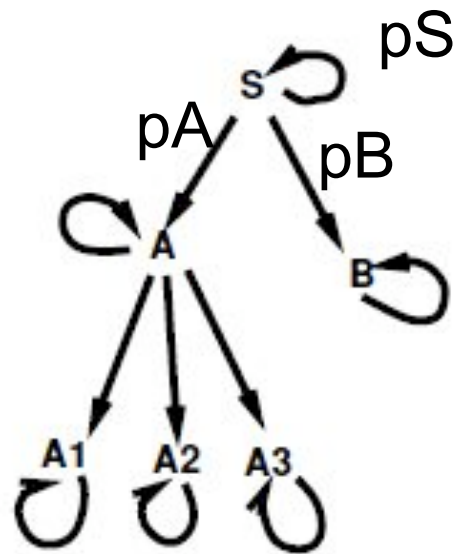


(both types are necessary)

Interaction works as bifurcation parameter for intracellular dynamics: self-consistency between intracellular dynamics and distribution of each cell type (Nakajima, 2007)

Hierarchical differentiation from  
'stem cell'; by taking initially  
dynamics with instability  
(e.g., chaotic)

stem cell as Milnor attractor?



probability depends on # distrib. of cell types  
with prob.  $pA$  for  $S \rightarrow A$

if  $\#(A)$  decreases then  $pA$  increases:

**STABILITY**

## Summary: Consistency Principle for Biology

(1) replication of molecules and cells : Universal Laws

(2) genetic and phenotypic changes

→ Phenotypic Fluctuation  $\propto$  Evolution Speed

→ Relation between

(isogenic)phenotype fluctuation vs

phenotype variation by mutation

Robustness to mutation and to developmental noise  
are linked

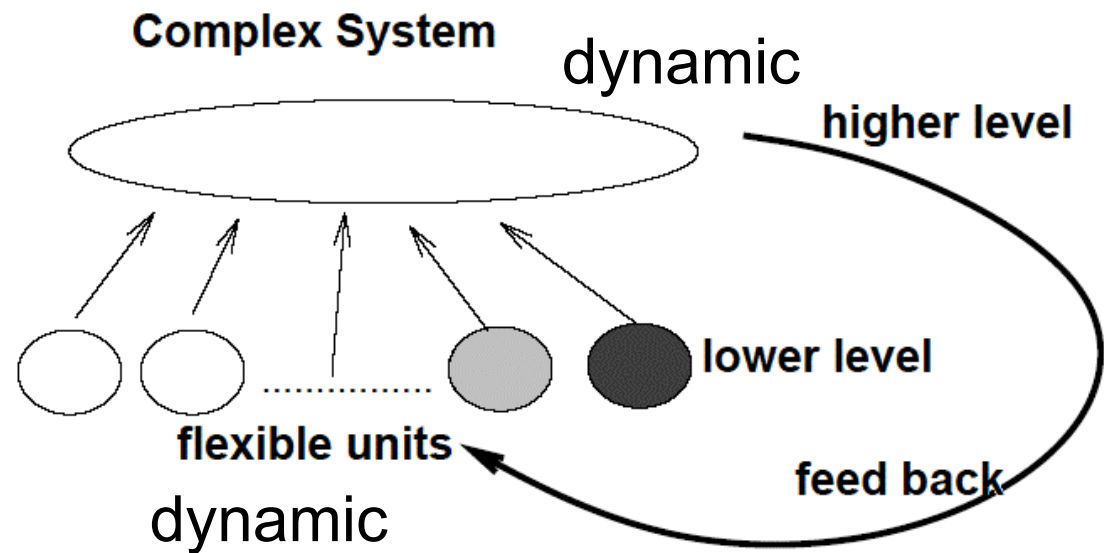
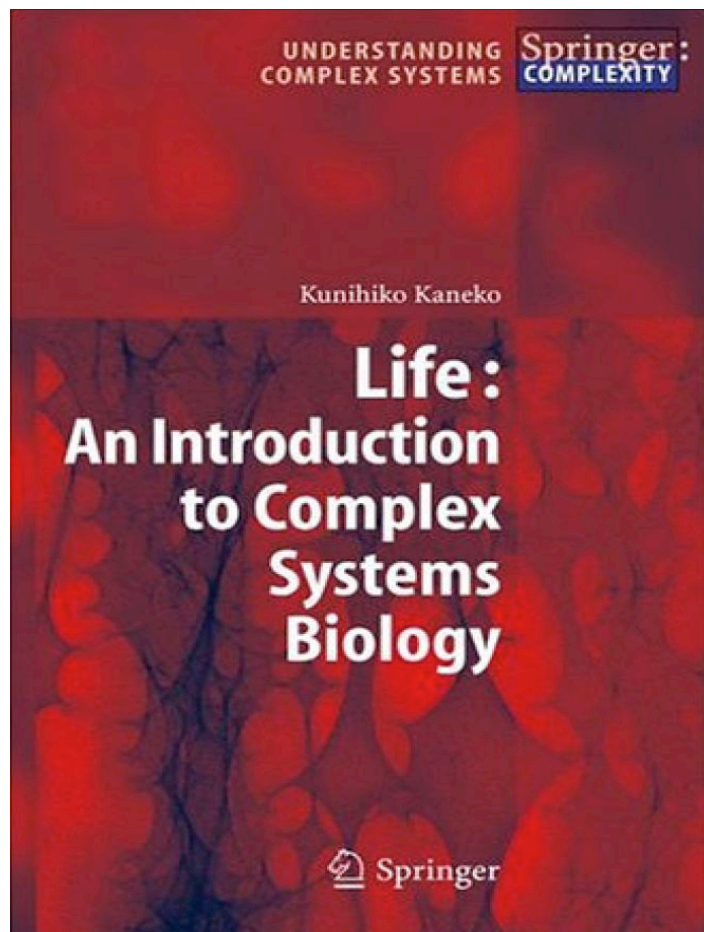
(3) adaptation of internal cellular state and growth

Growth system → general adaptation by noise

(4) replication of cells and cell ensembles

\*differentiation from stem cell, developmental robustness

Consistency Principle  
for stable state but  
for innovation,  
breakdown  
of consistency  
→ Chaotic Itinerancy



## Collaborators

**Chikara Furusawa**

Katsuhiko Sato

**experiment**

**Tetsuya Yomo**

**Yochiro Ito**

**Akiko Kashiwagi**

Most papers available at

<http://chaos.c.u-tokyo.ac.jp>